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PHARMACEUTICAL COMPOSITIONS COMPRISING POLYNUCLEOTIDES ENCODING A RAF PROTEIN

The present invention relates to pharmaceutical compositions comprising polynucleotides encoding a Raf protein, vectors, host cells, polypeptides encoded by said polynucleotides and agonists or antagonists thereof. The invention furthermore relates to uses of such pharmaceutical compositions for preventing or treating pathological conditions in which endothelial cells are involved or affected. Furthermore, the invention relates to methods for screening compounds acting as agonists or antagonists as well as to diagnostic compositions and methods.

Endothelial cells form the inner wall surface of blood vessels. The presence of endothelium in all tissues, and its central role in numerous vital functions such as tissue survival and blood clotting, places the vascular system at the core of many pathological conditions that affect the human population. Angiogenesis, the formation of new blood vessels, is essential for embryonic development, subsequent growth and tissue repair. This process is also essential in several pathological conditions, such as neoplasias. Generally, endothelial cells play a critical role in diseases such as cancer, stroke, cardiac infarction, circulatory problems, tissue injury, diabetic retinopathy, psoriasis, inflammation and atherosclerosis.

However, to date the molecular mechanisms that take place in the course of pathological conditions where endothelial cells are involved or affected are still poorly understood. A more detailed knowledge of the signal transduction and gene regulation pathways within endothelial cells would be promising for identifying or generating pharmacologically valuable compounds that may specifically influence disease states localized in endothelial cells. In this regard, it would be favorable to have access to molecules which are capable of effectively down or upregulating the activity of genes that are known to be involved in pathological conditions, as for example those outlined above.

Thus, the technical problem underlying the present invention is to provide means and methods for preventing or treating pathological conditions in which endothelial cells are involved or affected.

This technical problem is solved by the provision of the embodiments as characterized in the claims.

Accordingly, the present invention relates to pharmaceutical compositions comprising a compound selected from the group consisting of

- (i) polynucleotides encoding a Raf protein selected from the group consisting of
 - (a) polynucleotides encoding a polypeptide comprising the amino acid sequence of any one of SEQ ID NOs: 2, 4 and 6;
 - (b) polynucleotides comprising the coding region of the nucleotide sequence of any one of SEQ ID NOs: 1, 3 and 5;
 - (c) polynucleotides encoding a fragment of a polypeptide encoded by a polynucleotide of (a) or (b); and
 - (d) polynucleotides the complementary strand of which hybridizes with a polynucleotide of any one of (a) to (c) and encode a polynucleotide having Raf activity;
 - (ii) vectors comprising and capable of expressing said polynucleotide;
 - (iii) host cells genetically engineered with said polynucleotide or said vector;
 - (iv) polypeptides encoded by said polynucleotide; and
 - (v) agonists of a polypeptide encoded by said polynucleotide;
- and optionally a pharmaceutically acceptable carrier.

The present invention relates in a further embodiment to pharmaceutical compositions comprising a compound which is an antagonist of the polypeptide encoded by a polynucleotide as defined above and optionally a pharmaceutically acceptable carrier. The term "nucleic acid molecule" and "polynucleotide" are used interchangeably herein. The term "and/or" wherever used herein includes the meaning of "and", "or" and "any other combination of the elements connected by said term".

The invention, thus, relates in one aspect to pharmaceutical compositions that are based on a protein that belongs to the Raf protein kinase family and to polynucleotide encoding such proteins as identified above. The gene family of Raf protein kinases comprises the genes encoding the protein kinases A-Raf, B-Raf and C-Raf, from which the amino acid sequences of the human representatives is shown in the SEQ ID NOs: 2, 4 and 6, respectively. Corresponding cDNA sequences encoding these human A-Raf, B-Raf and C-Raf proteins are shown under SEQ ID NOs: 1, 3 and 5, respectively. The Raf proteins are protein kinases that are involved in numerous signal transduction cascades in eukaryotic cells. As a preferred embodiment, the present invention relates to pharmaceutical compositions, wherein the above mentioned Raf protein is B-Raf.

The present invention is based on the surprising finding that in a mouse embryonic endothelial progenitor cell (eEPC) line in which the gene encoding B-Raf is inactivated, in the following also referred to as B-Raf KO (knock out) or null cells, the expression profile of a large set of genes is altered compared to the corresponding wild-type (wt) cells. In the experiments which are described in detail in the appended Example 1, it could be shown that of the approximately 1500 genes so far studied about 5% to 10% are up or down regulated on the transcriptional level in the B-Raf KO cells (see Table 1). If one extrapolates from these results to the ten-thousands of genes that are normally expressed in wild-type embryonic endothelial cells, it is reasonable to anticipate that several hundred to several thousand cellular genes show an altered gene expression in B-Raf KO cells and are therefore influenced by signaling pathways under control of B-Raf.

In subsequent experiments, these results obtained with B-Raf null cells were confirmed for the situation in vivo by comparing gene expression profiles between wt and B-Raf KO embryos (Example 2). The results show that genes, whose expression changed in the B-Raf null cells, were affected during embryonic vascular development, i.e. they were also deregulated.

Further evidence that B-Raf is essential for protein regulation in endothelial cells is provided by the fact that cAMP can induce the activation of the B-Raf mediated MAP kinase pathway in eEPCs as shown in Example 3. These results suggest that the effects of a number of diverse extracellular signals that modulate intracellular cAMP

levels in endothelial cells, and thus lead to MAPK pathway activation, can in turn be modulated by agents that affect B-Raf activity.

In addition, the electron microscopy data shown in Example 4 and Figure 4, reveal B-Raf's importance for the proper establishment of the endothelial/peri-endothelial environment. This and the similarity to the angiogenin 1 (Ang1) KO mice phenotype observed, which points at a position of B-Raf downstream from tie-2 in the signaling cascade, substantiates the role of B-Raf in angiogenesis, wound healing and endothelial cell migration.

These data might explain the fact that the B-Raf KO genotype is lethal for homozygous knock out mice already in the mid-gestation phase and that in such mice endothelial cells are extensively affected by apoptosis (Wojnowski, *Nature Genetics* 16 (1997), 293-297).

This central role of B-Raf in the gene regulation of endothelial cells was unexpected because B-Raf protein is not endothelial-specific but is expressed in many cell types (Bernier, *J. Biol. Chem.* 270 (1995), 23381-23389) and, in particular, rather extensively in neuronal tissues (Jaiswal et al., *Mol. Cell. Biol.* 14 (1994), 6944-6953).

The consequences of the B-Raf pathway impairment in endothelial cells were previously not known. The above-mentioned results demonstrate for the first time that gene expression of a large number of genes is altered and that the affected genes belong to molecular pathways with crucial roles in endothelial cell functions such as angiogenesis, thrombosis, atherosclerosis and inflammation.

These data lead to two major conclusions: (a) the results obtained in B-Raf null cells are not an artifact of the cell isolation procedure because of their reproducibility in B-Raf KO embryos, and (b) the documented changes in gene expression profiles are a direct consequence of the lack of B-Raf protein in endothelial cells. Consequently, B-Raf has a central regulatory role in vascular system diseases that affect endothelial cells.

It is therefore conceivable to use pharmaceutical compositions suitable for modulation of B-Raf activity in therapeutic approaches for pathological conditions in which endothelial cells are involved or affected, such as tumor-induced angiogenesis, diabetic retinopathy, wound healing, psoriasis, inflammation, stroke, myocardial infarction, atherosclerosis and arterial restenosis.

The therapeutic uses conceivable for compounds modulating B-Raf activity in endothelial cells are also envisaged for A-Raf and C-Raf because biochemical analyses have shown that the distinct functions of the three types of kinases overlap to a high degree (Pritchard, Mol. Cell. Biol. 15 (1995), 6430-6442). Moreover, genetic and phenotypic analysis in single gene KO mice and in double B-Raf and C-Raf KO mice show that the functions for example of B-Raf and C-Raf are overlapping but not redundant (Wojnowski, Mechanisms of Development 91 (2000), 97-104).

In one embodiment, the pharmaceutical compositions of the invention comprise a compound which is a polynucleotide encoding a Raf protein.

The term "Raf protein" refers to a protein encoded by a member of the Raf protein kinase gene family. In particular, this gene family encodes three different types of protein kinases, A-Raf, B-Raf and C-Raf. As a common feature, these proteins are part of various signal transduction pathways. Structure, activity and functions of the members of the Raf kinase gene family are described in detail in the prior art as for instance reviewed in Morrison and Cutler (Current Opinion in Cell Biology 9 (1997), 174-179). A common feature of the Raf family of protein kinases is that they all share three highly conserved regions called CR1, CR2 and CR3. CR1 is rich in cysteine residues while CR2 contains many serines and threonines. The CR3 domain contains the kinase activity. In accordance with the sequence comparisons described in Sithanandam (Oncogene 5 (1999), 1775-1780), CR1 and CR3 are highly conserved among all three types of protein kinases, preferably having a homology, that is to say amino acid sequence identity of more than 90%, more preferably of more than 93%. CR2, in contrast thereto, is highly conserved only between A-Raf and C-Raf, while it is less conserved in B-Raf, having a homology of less than 60%, preferably of less than 50% compared to CR2 of A-Raf and C-Raf. The naturally occurring Raf proteins feature considerable size differences. On average, B-Raf proteins have a molecular weight of about 90 kDa and A-Raf and C-Raf of about 70 kDa. Correspondingly, the amino acid sequence of A-Raf and C-Raf is at least 100 residues shorter than that of B-Raf. Examples of proteins that are capable of activating Raf proteins include ras and Protein Kinase A (PKA). Another kinase, Rap1, is capable of phosphorylating B-Raf while it does not phosphorylate C-Raf. The kinase activity of Raf proteins leads to phosphorylation of specific substrate proteins, thereby leading to an activation or

inactivation, preferably to an activation, of the substrate protein. Examples of pathways that Rafs are involved in are the MAP kinase pathway and the MEK/ERK pathway (Avruch, Trends Biochem. Sci. 19 (1994), 279-283). Specifically B-Raf is capable of phosphorylating the proteins MEK 1 and 2 (Heidecker, Mol. Cell. Biol. 10 (1990), 2503-2512; Reuter, J. Biol. Chem. 270 (1995), 7644-7655). Suitable assays for determining Raf protein activity are described in the literature, for instance in Reuter (J. Biol. Chem. 270 (1995), 7644-7655), Jaiswal (Mol. Cell. Biol. 14 (1994), 6944-6953) and Voltek (Cell 17 (1993), 205-214; Wang, Cell 87 (1996), 629-638). A specific example of a kinase assay to monitor Raf kinase activity is as follows: Raf kinases are purified from eukaryotic cells following immunoprecipitation with Anti-Raf antibodies. The collected complexes are washed in salt/HEPES buffer and then incubated with baculovirus-produced MEK protein in the presence of radioactive gamma ATP. Following incubation for 30 min at 30°C, the reaction products (phosphorylated MEK) are resolved in 10% polyacrylamide gels, visualized by autoradiography and counted using a Phosphoimager. Suitable assays to distinguish A-, B- and C-Raf activity may use differences regarding potency in activating MEK. For instance, according to an in vitro assay described by Pritchard (Mol. Cell. Biol. 15 (1995), 6430-6442), B-Raf phosphorylating activity is about 10 times stronger than that of C-Raf and about 500 times stronger than that of A-Raf. According to another approach, specific antibodies against A-, B- and C-Raf can be applied to monitor their phosphorylation status (Reuter, J. Biol. Chem. 270 (1995), 7644-7655).

Preferably, the polynucleotide encodes a Raf protein comprising the amino acid sequence of the human A-Raf, B-Raf or C-Raf protein, i.e. the sequences shown in SEQ ID NOs: 2, 4 and 6, respectively. Such polypeptides are preferably encoded by the polynucleotides comprising the coding region of the nucleotide sequence shown in SEQ ID NOs: 1, 3 and 5, respectively, or are retrievable from the GeneBank/EMBL data base entries having the accession Nos. X04790, M95712 and X03484, respectively. As a preferred embodiment, said polynucleotides encode a polypeptide comprising the amino acid sequence of SEQ ID NO: 4 or comprises the coding region of the nucleotide sequence of SEQ ID NO: 3.

The polynucleotide contained in the pharmaceutical composition of the invention may as well encode a fragment or an epitope-bearing portion of a polypeptide as

described above. Preferably, such fragments have the activity of a Raf protein. In particular, the length of such a fragment is not below 10 amino acid residues, preferably not below 20, more preferably not below 50 and most preferably not below 100 amino acid residues.

In another embodiment the polynucleotide contained in the pharmaceutical composition is a polynucleotide the complementary strand of which hybridizes with one of the above-described polynucleotides and which encodes a Raf protein.

Also included in this context are polynucleotides which encode a protein, which has a homology, that is to say a sequence identity, of at least 30%, preferably of at least 40%, more preferably of at least 50%, even more preferably of at least 60% and particularly preferred of at least 70%, especially preferred of at least 80% and even more preferred of at least 90% to the entire amino acid sequence as indicated in SEQ ID NO: 2, 4, or 6 the protein being a Raf protein.

Hybridizing polynucleotides also encompass polynucleotides which encode a Raf protein and the nucleotide sequences of which have a homology, that is to say a sequence identity, of at least 40%, preferably of at least 50%, more preferably of at least 60%, even more preferably of at least 70%, in particular of at least 80%, especially preferred of at least 90%, in particular of at least 95% and even more preferred of at least 99% when compared to the coding region of the sequence shown in SEQ ID NO:1, 3 or 5.

In the context of the present invention the term "hybridization" means hybridization under conventional hybridization conditions, preferably under stringent conditions, as for instance described in Sambrook et al., Molecular Cloning, A Laboratory Manual, 2nd edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY. In an especially preferred embodiment the term "hybridization" means that hybridization occurs under the following conditions:

Hybridization buffer: 2 x SSC; 10 x Denhardt solution (Fikoll 400 + PEG + BSA; ratio 1:1:1); 0.1% SDS; 5 mM EDTA; 50 mM Na₂HPO₄;

250 µg/ml of herring sperm DNA; 50 µg/ml of tRNA;
or
0.25 M of sodium phosphate buffer, pH 7.2;
1 mM EDTA
7% SDS
Hybridization temperature T = 60°C
Washing buffer: 2 x SSC; 0.1% SDS
Washing temperature T = 60°C.

Polynucleotides which hybridize with the polynucleotides as specified above can, in principle, encode a Raf protein from any organism expressing such proteins or can encode modified versions thereof.

Polynucleotides which hybridize with the polynucleotides as specified above can for instance be isolated from genomic libraries or cDNA libraries of bacteria, fungi, plants or animals. Preferably, such molecules are from animal origin, e.g., from vertebrate or evertbrate animals, more preferably from mammal origin and particularly preferred from human origin. It is furthermore preferred that the polynucleotide used in the pharmaceutical composition of the invention is isolated from the same species as the species to which the pharmaceutical composition is to be administered. In this regard, the species may be any one to which administration of the pharmaceutical composition is useful, for example to vertebrates, preferably to mammals. Preferred mammals are rodents (e.g. mouse or rat), working animals (e.g. cow, horse, pig), other domestic animals (e.g. dog, cat), animals commonly used in animal experiments (in addition to other already mentioned species, e.g., rabbit, monkey), and most preferred are humans. As an alternative to isolate said polynucleotides from natural sources, they can be prepared by genetic engineering or chemical synthesis.

Polynucleotides useful for preparing the pharmaceutical composition of the invention may be identified and isolated by using the above-described polynucleotides or parts of these polynucleotides or reverse complements of these polynucleotides, for instance by hybridization according to standard methods (see for instance Sambrook et al., 1989, Molecular Cloning. A Laboratory Manual, 2nd edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY).

Nucleic acid molecules comprising the same or substantially the same nucleotide sequence as indicated in SEQ ID NO: 1, 3 or 5 or fragments thereof can, for instance, be used as hybridization probes. Such fragments used as hybridization probes can also be synthetic fragments which are prepared by usual synthesis techniques.

The polynucleotides hybridizing with the above-described polynucleotides also comprise fragments, derivatives and allelic variants of said above-described polynucleotides encoding a Raf protein. Herein, fragments are understood to mean parts of the polynucleotides which are long enough to encode the described protein, preferably showing the biological activity of a Raf protein described above, e.g. being capable of activating MEK protein and/or the MEK/ERK signaling cascade. In this context, the term derivative means that the sequences of these molecules differ from the sequences of the above-described polynucleotides in one or more positions and show a high degree of homology to these sequences. In this context, homology means a sequence identity of at least 40%, in particular an identity of at least 60%, preferably of more than 65%, even more preferably of at least 70%, in particular of at least 80%, more preferably of at least 90% and particularly preferred of more than 95%. Deviations from the above-described polynucleotides may have been produced, e.g., by deletion, substitution, insertion and/or recombination.

Preferably, the degree of homology is determined by comparing the respective sequence with the nucleotide sequence of the coding region of SEQ ID NO: 1, 3 or 5. When the sequences which are compared do not have the same length, the degree of homology preferably refers to the percentage of nucleotide residues in the shorter sequence which are identical to nucleotide residues in the longer sequence. The degree of homology can be determined conventionally using known computer programs such as the DNASTAR program with the ClustalW analysis. This program can be obtained from DNASTAR, Inc., 1228 South Park Street, Madison, WI 53715 or from DNASTAR, Ltd., Abacus House, West Ealing, London W13 0AS UK (support@dnastar.com) and is accessible at the server of the EMBL outstation.

When using the Clustal analysis method to determine whether a particular sequence is, for instance, 80% identical to a reference sequence the settings are preferably as follows: Matrix: blosum 30; Open gap penalty: 10.0; Extend gap penalty: 0.05; Delay

divergent: 40; Gap separation distance: 8 for comparisons of amino acid sequences. For nucleotide sequence comparisons, the Extend gap penalty is preferably set to 5.0.

Furthermore, homology means preferably that the encoded protein displays a sequence identity of at least 30%, more preferably of at least 40%, even more preferably of at least 50%, in particular of at least 60%, particularly preferred of at least 70%, especially preferred of at least 80% and even more preferred of at least 90% to the amino acid sequence depicted under SEQ ID NO: 2, 4 or 6.

Preferably, the degree of homology of the hybridizing polynucleotides is calculated over the complete length of its coding sequence. It is furthermore preferred that such a hybridizing nucleic acid molecule, and in particular the coding sequence comprised therein, has a length of at least 200 nucleotides, preferably at least 400 nucleotides, more preferably of at least 600 nucleotides, even more preferably of at least 800 nucleotides, particularly preferred of at least 1000 nucleotides and most preferably of at least 1500 nucleotides.

Preferably, sequences hybridizing to a polynucleotide as described above comprise a region of homology of at least 90%, preferably of at least 93%, more preferably of at least 95%, still more preferably of at least 98% and particularly preferred of at least 99% identity to said polynucleotide, wherein this region of homology has a length of at least 300 nucleotides, more preferably of at least 500 nucleotides, even more preferably of at least 750 nucleotides, particularly preferred of at least 1000 nucleotides and most preferably of at least 1500 nucleotides.

Homology, moreover, means that there is a functional and/or structural equivalence between the corresponding polynucleotides or proteins encoded thereby. Polynucleotides which are homologous to the above-described molecules and represent derivatives of these molecules are normally variations of these molecules which represent modifications having the same biological function. They may be either naturally occurring variations, for instance sequences from other allelic variants, varieties, species, etc., or mutations, and said mutations may have formed naturally or may have been produced by deliberate mutagenesis. Furthermore, the variations may be synthetically produced sequences. The allelic variants may be naturally occurring variants or synthetically produced variants or variants produced by recombinant DNA techniques.

The proteins encoded by the different variants of the above-described polynucleotides possess certain characteristics they have in common. These include for instance biological activity, molecular weight, immunological reactivity, conformation, etc., and physical properties, such as for instance the migration behavior in gel electrophoreses, chromatographic behavior, sedimentation coefficients, solubility, spectroscopic properties, stability, pH optimum, temperature optimum etc.

The biological activity of the Raf protein, in particular the capacity to phosphorylate MEK or can be tested as described above, e.g. by an assay as described in Reuter (J. Biol. Chem. 270 (1995), 7644-7655).

The polynucleotide contained in the pharmaceutical composition of the invention may also comprise in addition to the coding sequence for a Raf protein, as for example the polypeptide having the amino acid sequence of SEQ ID NO: 2, 4 or 6 or a polypeptide homologous thereto, further coding or non-coding nucleotide sequences. Thus, for instance, the polynucleotide may encode one of the above-described polypeptides which is fused to a marker sequence, such as a peptide, which facilitates purification of the fused polypeptide. The marker sequence may for example be a hexa-histidine peptide, such as the tag contained in a pQE vector (Qiagen, Inc.) which provides for convenient purification of the fusion protein. Another suitable marker sequence may be the HA tag which corresponds to an epitope derived from influenza hemagglutinin protein (Wilson, Cell 37 (1984), 767). A further example is the coding sequence of glutathione-S-transferase (GST) which, apart from providing a purification tag, enhances protein stability, for instance, in bacterial expression systems.

The polynucleotide contained in the pharmaceutical composition of the invention can be any type of polynucleotide, e.g. DNA molecules or RNA molecules or combinations thereof. These polynucleotides can be obtained for instance from natural sources or may be produced synthetically or by recombinant techniques, such as PCR. Such polynucleotides may comprise any modification thereof that is known in the state of the art (see, e.g., US 5525711, US 4711955, US 5792608 or EP 302175 for examples of modifications). Such polynucleotides may be single- or

double-stranded, linear or circular, without any size limitation. For instance, the polynucleotide(s) may be genomic DNA, cDNA, mRNA etc.

The polynucleotides described hereinabove allow to prepare host cells as well as to produce recombinantly proteins having Raf activity of high purity and/or in sufficient quantities for preparing pharmaceutical compositions comprising any of these compounds.

Another compound that may be comprised in a pharmaceutical composition of the invention is a vector comprising and capable of expressing a polynucleotide as described above. Such a vector can be an expression vector and/or a gene delivery vector. Expression vectors are in this context meant for use in ex vivo gene therapy techniques, i.e. suitable host cells are transfected outside the body and then administered to the subject. Gene delivery vectors are referred to herein as vectors suited for in vivo gene therapeutic applications, i.e. the vector is directly administered to the subject, either systemically or locally. The vector referred to herein may only consist of nucleic acid or may be complexed with additional compounds that enhance, for instance, transfer into the target cell, targeting, stability and/or bioavailability, e.g. in the circulatory system. Examples of such additional compounds are lipidic substances, polycations, membrane-disruptive peptides or other compounds, antibodies or fragments thereof or receptor-binding molecules specifically recognizing the target cell, etc. Expression or gene delivery vectors may preferably be derived from viruses such as retroviruses, vaccinia virus, adeno-associated virus, herpes viruses or bovine papilloma virus, and may be used for delivery into a targeted cell population, e.g. into endothelial cells. Methods which are well known to those skilled in the art can be used to construct recombinant expression or gene delivery vectors; see, for example, the techniques described in Sambrook, *Molecular Cloning A Laboratory Manual*, Cold Spring Harbor Laboratory (1989) N.Y. and Ausubel, *Current Protocols in Molecular Biology*, Green Publishing Associates and Wiley Interscience, N.Y. (1989). Alternatively, the vectors can be reconstituted into liposomes for delivery to target cells. The vectors containing the above-described polynucleotides can be transferred into a host cell by well-known methods, which vary depending on the type of cellular host. For example, calcium

chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cellular hosts; see Sambrook, loc. cit.

The vectors referred to herein may comprise further genes such as marker genes which allow for the selection of said vector in a suitable host cell and under suitable conditions. Preferably, the polynucleotide comprised in the vector is operatively linked to expression control sequences allowing expression in prokaryotic or eukaryotic cells. Expression of said polynucleotide comprises transcription of the polynucleotide into a translatable mRNA. Regulatory elements ensuring expression in eukaryotic cells, preferably mammalian cells, are well known to those skilled in the art. They usually comprise regulatory sequences ensuring initiation of transcription and optionally poly-A signals ensuring termination of transcription and stabilization of the transcript, and/or an intron further enhancing expression of said polynucleotide. Additional regulatory elements may include transcriptional as well as translational enhancers, and/or naturally-associated or heterologous promoter regions. Possible regulatory elements permitting expression in prokaryotic host cells comprise, e.g., the PL, lac, trp or tac promoter in *E. coli*, and examples for regulatory elements permitting expression in eukaryotic host cells are the AOX1 or GAL1 promoter in yeast or the CMV-, SV40-, RSV-promoter (Rous sarcoma virus), CMV-enhancer, SV40-enhancer or a globin intron in mammalian or other animal cells. Beside elements which are responsible for the initiation of transcription such regulatory elements may also comprise transcription termination signals, such as the SV40-poly-A site or the tk-poly-A site, downstream of the polynucleotide. Furthermore, depending on the expression system used leader sequences capable of directing the polypeptide to a cellular compartment or secreting it into the extracellular space may be added to the coding sequence of the polynucleotide and are well known in the art. The leader sequence(s) is (are) assembled in appropriate phase with translation, initiation and termination sequences, and preferably, a leader sequence capable of directing secretion of the translated protein, or a portion thereof, into the periplasmic space or extracellular space. Optionally, the heterologous sequence can encode a fusion protein including an C- or N-terminal identification peptide imparting desired characteristics, e.g., stabilization or simplified purification of expressed recombinant product. In this context, suitable expression vectors are known in the art such as

Okayama-Berg cDNA expression vector pcDV1 (Pharmacia), pCDM8, pRc/CMV, pcDNA1, pcDNA3 (In-vitrogen), or pSPORT1 (GIBCO BRL).

Preferably, the expression control sequences will be eukaryotic promoter systems in vectors capable of transforming or transfecting eukaryotic host cells, but control sequences for prokaryotic hosts may also be used.

As mentioned above, the vector described herein is for use in gene therapy, which is based on introducing therapeutic genes into cells by ex-vivo or in-vivo techniques. Suitable vectors and methods for ex-vivo or in-vivo gene therapy are described in the literature and are known to the person skilled in the art; see, e.g., Giordano, *Nature Medicine* 2 (1996), 534-539; Schaper, *Circ. Res.* 79 (1996), 911-919; Anderson, *Science* 256 (1992), 808-813; Isner, *Lancet* 348 (1996), 370-374; Muhlhauser, *Circ. Res.* 77 (1995), 1077-1086; Wang, *Nature Medicine* 2 (1996), 714-716; WO 94/29469; WO 97/00957 or Schaper, *Current Opinion in Biotechnology* 7 (1996), 635-640, and references cited therein. The vectors comprised in the pharmaceutical composition of the invention may be designed for direct introduction or for introduction via liposomes or viral vectors (e.g. adenoviral, retroviral) into the cell. Preferably, said cell is a germ line cell, embryonic cell or egg cell or a cell derived therefrom, most preferably said cell is a stem cell.

In a further embodiment, the pharmaceutical compositions of the invention comprise host cells genetically engineered with the above-described polynucleotides or vectors.

Said host cell may in principle be a prokaryotic or eukaryotic cell. Said polynucleotide or vector which is present in the host cell may either be integrated into the genome of the host cell or it may be maintained extrachromosomally. The term "prokaryotic" is meant to include all bacteria which can be transformed or transfected with said polynucleotides or vectors. Prokaryotic hosts may include gram negative as well as gram positive bacteria such as, for example, *E. coli*, *S. typhimurium*, *Serratia marcescens* and *Bacillus subtilis*. The term "eukaryotic" is meant to include fungal higher plant, insect and preferably mammalian cells. Preferred fungal cells are yeast cells, for example, those of the genus *Saccharomyces*, in particular those of the species *S. cerevisiae*. Among the mammalian cells human cells are most preferred.

The transformation or transfection of the host cell with a polynucleotide or vector as described above can be carried out by standard methods, as for instance described

in Sambrook et al., (Molecular Cloning: A Laboratory Manual, 2nd edition (1989) Cold Spring Harbor Press, New York; Methods in Yeast Genetics, A Laboratory Course Manual, Cold Spring Harbor Laboratory Press, 1990).

The herein described host cells may be of therapeutic value in ex-vivo gene therapy, in particular by targeting them to the endothelium using a suitable targeting technique. Advantageously, such host cells are endothelial cells or progenitor cells thereof. For example, transplantation of genetically modified cells can be used for therapeutic purposes by providing endothelial cells with desired traits such as resistance to angiogenic signals or thromboresistance.

Furthermore, the pharmaceutical composition of the invention may comprise a polypeptide encoded by the above-described polynucleotide.

This polypeptide may, e.g., be a naturally purified product, or a product of chemical synthetic procedures, or produced by recombinant techniques from a prokaryotic or eukaryotic host (for example, by bacterial, yeast, higher plant, insect and mammalian cells in culture).

Provision of the polypeptide via recombinant expression of one of the above-described polynucleotides is preferred. An overview of different expression systems is for instance contained in Methods in Enzymology 153 (1987), 385-516, in Bitter et al. (Methods in Enzymology 153 (1987), 516-544) and in Sawers et al. (Applied Microbiology and Biotechnology 46 (1996), 1-9), Billman-Jacobe (Current Opinion in Biotechnology 7 (1996), 500-4), Hockney (Trends in Biotechnology 12 (1994), 456-463), Griffiths et al., (Methods in Molecular Biology 75 (1997), 427-440). An overview of yeast expression systems is for instance given by Hensing et al. (Antonie van Leeuwenhoek 67 (1995), 261-279), Bussineau et al. (Developments in Biological Standardization 83 (1994), 13-19), Gellissen et al. (Antonie van Leeuwenhoek 62 (1992), 79-93, Fleer (Current Opinion in Biotechnology 3 (1992), 486-496), Vedvick (Current Opinion in Biotechnology 2 (1991), 742-745) and Buckholz (Bio/Technology 9 (1991), 1067-1072).

Expression vectors have been widely described in the literature. As a rule, they contain not only a selection marker gene and a replication-origin ensuring replication in the host selected, but also a bacterial or viral promoter, and in most cases a termination signal for transcription. Between the promoter and the termination signal

there is in general at least one restriction site or a polylinker which enables the insertion of a coding DNA sequence. The DNA sequence naturally controlling the transcription of the corresponding gene can be used as the promoter sequence, if it is active in the selected host organism. However, this sequence can also be exchanged for other promoter sequences. It is possible to use promoters ensuring constitutive expression of the gene and inducible promoters which permit a deliberate control of the expression of the gene. Bacterial and viral promoter sequences possessing these properties are described in detail in the literature. Regulatory sequences for the expression in microorganisms (for instance *E. coli*, *S. cerevisiae*) are sufficiently described in the literature. Promoters permitting a particularly high expression of a downstream sequence are for instance the T7 promoter (Studier et al., Methods in Enzymology 185 (1990), 60-89), lacUV5, trp, trp-lacUV5 (DeBoer et al., in Rodriguez and Chamberlin (Eds), Promoters, Structure and Function; Praeger, New York, (1982), 462-481; DeBoer et al., Proc. Natl. Acad. Sci. USA (1983), 21-25), lp1, rac (Boros et al., Gene 42 (1986), 97-100). Inducible promoters are preferably used for the synthesis of proteins. These promoters often lead to higher protein yields than do constitutive promoters. In order to obtain an optimum amount of protein, a two-stage process is often used. First, the host cells are cultured under optimum conditions up to a relatively high cell density. In the second step, transcription is induced depending on the type of promoter used. In this regard, a tac promoter is particularly suitable which can be induced by lactose or IPTG (=isopropyl- β -D-thiogalactopyranoside) (deBoer et al., Proc. Natl. Acad. Sci. USA 80 (1983), 21-25). Termination signals for transcription are also described in the literature.

Transformation or transfection of suitable host cells can be carried out according to one of the methods mentioned above. The host cell is cultured in nutrient media meeting the requirements of the particular host cell used, in particular in respect of the pH value, temperature, salt concentration, aeration, antibiotics, vitamins, trace elements etc. The polypeptide having Raf activity can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography.

Protein refolding steps can be used, as necessary, in completing configuration of the protein. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps.

Depending upon the host employed in a recombinant production procedure, the polypeptide may be glycosylated or may be non-glycosylated. The polypeptide may also include an initial methionine amino acid residue. The polypeptide may be further modified to contain additional chemical moieties not normally part of a naturally occurring protein. Those derivatized moieties may, e.g., improve the stability, solubility, the biological half life or absorption of the polypeptide. The moieties may also reduce or eliminate any undesirable side effects of the polypeptide and the like. An overview for these moieties can be found, e.g., in Remington's Pharmaceutical Sciences (18th ed., Mack Publishing Co., Easton, PA (1990)). Polyethylene glycol (PEG) is an example for such a chemical moiety which has been used for the preparation of therapeutic proteins. The attachment of PEG to proteins has been shown to protect them against proteolysis (Sada et al., J. Fermentation Bioengineering 71 (1991), 137-139). Various methods are available for the attachment of certain PEG moieties to proteins (for review see: Abuchowski et al., in "Enzymes as Drugs"; Holcerberg and Roberts, eds. (1981), 367-383). Generally, PEG molecules are connected to the protein via a reactive group found on the protein. Amino groups, e.g. on lysines or the amino terminus of the protein are convenient for this attachment among others.

The polypeptides comprised in the pharmaceutical compositions of the present invention can comprise a further domain, said domain being linked by covalent or non-covalent bonds. The linkage can be based on genetic fusion according to the methods known in the art and described above or can be performed by, e.g., chemical cross-linking as described in, e.g., WO 94/04686. The additional domain present in the fusion protein comprising the polypeptide employed in accordance with the invention may preferably be linked by a flexible linker, advantageously a polypeptide linker, wherein said polypeptide linker comprises plural, hydrophilic, peptide-bonded amino acids of a length sufficient to span the distance between the C-terminal end of said further domain and the N-terminal end of the polypeptide or vice versa. The above described fusion protein may further comprise a cleavable linker or cleavage site for proteinases.

Furthermore, said further domain may be of a predefined specificity or function. In this context, it is understood that the polypeptides present in the pharmaceutical composition according to the invention may be further modified by conventional methods known in the art. This allows for the construction of fusion proteins comprising the polypeptide of the invention and other functional amino acid sequences, e.g., nuclear localization signals, transactivating domains, DNA-binding domains, hormone-binding domains, protein tags (GST, GFP, h-myc peptide, FLAG, HA peptide) which may be derived from heterologous proteins. Thus, administration of the composition of the invention can utilize unlabeled as well as labeled polypeptides.

The pharmaceutical compositions of the invention may also comprise compounds that have an agonistic effect on the polypeptide having Raf activity as described above.

Preferably, said agonist compound can be identified by a method for screening such compounds comprising the steps of

- (a) measuring the activity of a polypeptide as described above in the presence and in the absence of the compound to be tested; and
- (b) identifying a compound acting as an agonist if the activity measured in step (a) is higher in the presence of the compound than in the absence of the compound.

Screening methods for identifying compounds that influence the activity of a given protein are well known in the art and can be taken from the literature. Preferably, such a screening is carried out in a high-throughput fashion with a degree of automation as high as possible. Candidate compounds may for instance be provided from libraries of chemical or biological substances that are routinely taken for such approaches and are known in the art. The activity assayed in step (a) is preferably the kinase activity of the Raf protein. The kinase activity may be measured as described above. Further activity assays are for instance described in Reuter (J. Biol. Chem. 270 (1995), 7644-7655), Jaiswal (Mol. Cell. Biol. 14 (1994), 6944-6953), Voltek (Cell 74 (1993), 205-214) and Wang (Cell 87 (1996), 629-638). For instance, such an activity assay can be carried out as described above, i.e. using

recombinantly produced MEK in the presence of radioactively labeled gamma ATP. One could furthermore use an apoptosis assay (e.g. the TUNEL ASSAY) or one could do RNA analysis for changes in gene expression based on the genes identified in the context of the present invention (see Table 1).

The present invention furthermore relates to pharmaceutical compositions comprising a compound which is an antagonist of the above-described polypeptide having Raf kinase activity.

Potential Raf protein antagonists include antibodies or fragments thereof or oligonucleotides which bind to the polypeptide and effectively reduce Raf kinase activity.

Antibodies useful as antagonists can be monoclonal or polyclonal and can be prepared according to methods well known in the art. The term "antibody" also comprises fragments of an antibody which still retain the binding specificity.

The polypeptide as described above, its fragments or other derivatives thereof, or cells expressing them can be used as an immunogen to produce antibodies thereto. In particular, also included are chimeric, single chain, and humanized antibodies, as well as Fab fragments, or the product of a Fab expression library. Various procedures known in the art may be used for the production of such antibodies and fragments.

Antibodies directed against a polypeptide as described above can be obtained, e.g., by direct injection of the protein into an animal or by administering the polypeptide to an animal, preferably a non-human animal. The antibody so obtained will then bind the protein itself. In this manner, even a sequence encoding only a fragment of the protein can be used to generate antibodies binding the whole native polypeptide.

Such antibodies can then, e.g., be used to isolate the protein from tissue expressing that polypeptide or to detect it in a probe. For the preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples for such techniques include the hybridoma technique (Köhler and Milstein, Nature 256 (1975), 495-497), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., Immunology Today 4 (1983), 72) and the EBV-hybridoma technique to produce human monoclonal antibodies

(Cole et al., Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc. (1985), 77-96). Techniques describing the production of single chain antibodies (e.g., U.S. Patent 4,946,778) can be adapted to produce single chain antibodies to immunogenic polypeptides as described above. Furthermore, transgenic mice may be used to express humanized antibodies directed against said immunogenic polypeptides.

Alternatively, a potential antagonist may be a mutant form of the polypeptide, preferably a dominant mutant form, which is for example inactive with regard to biological activity, e.g. to kinase activity. Such a mutant form could, for instance, bind to the natural Raf substrate protein but without activating it through phosphorylation. Alternatively or in addition, the mutant form could still have a phosphorylation site whereby phosphorylation of this site does not result in Raf activation. Such inactivated mutant forms of Raf proteins would be suited to block the natural Raf-mediated signal transduction pathway. Since protein structures of A-, B-, as well as of C-Raf proteins are well known in the art, the person skilled in the art is capable of designing such mutant forms from the corresponding wild-type Raf protein by way of amino acid deletion(s), substitution(s) and/or addition(s) in the amino acid sequence of the protein.

Another class of potential antagonist compounds comprises nucleic acid molecules that are capable of reducing Raf protein activity in a cell by way of intervening into gene expression of said protein, such as for example antisense, sense, ribozyme or co-suppression constructs. Antisense technology can be used to control gene expression through triple-helix formation or antisense DNA or RNA, whereby the inhibitory effect is based on specific binding of a nucleic acid molecule to DNA or RNA. For example, the 5' coding portion of a nucleic acid molecule encoding a Raf protein to be inhibited can be used to design an antisense RNA oligonucleotide, e.g., of from about 10 to 40 nucleotides in length. The antisense DNA or RNA oligonucleotide hybridizes to the mRNA in vivo and blocks translation of said mRNA and/or leads to destabilization of the mRNA molecule (Okano, J. Neurochem. 56 (1991), 560; Oligodeoxynucleotides as antisense inhibitors of gene expression, CRC Press, Boca Raton, FL, USA (1988)). For applying a triple-helix approach, a DNA

oligonucleotide can be designed to be complementary to a region of the gene encoding a Raf protein to be inhibited according to the principles laid down in the prior art (see for example Lee, Nucl. Acids Res. 6 (1979), 3073; Cooney, Science 241 (1988), 456; and Dervan, Science 251 (1991), 1360). Such a triple-helix forming oligonucleotide can then be used to prevent transcription of the specific gene. The oligonucleotides described above can also be delivered to target cells via a gene delivery vector as described above in order to express such molecules in vivo to inhibit gene expression of the respective Raf protein.

Examples for antisense molecules are oligonucleotides specifically hybridizing to a polynucleotide encoding a polypeptide having Raf activity. Such oligonucleotides have a length of preferably at least 10, in particular at least 15, and particularly preferably of at least 50 nucleotides. They are characterized in that they specifically hybridize to said polynucleotide, that is to say that they do not or only to a very minor extent hybridize to other nucleic acid sequences.

Likewise, RNA molecules with ribozyme activity which specifically cleave transcripts of a gene encoding a Raf protein or DNA molecules encoding such RNAs can be used. Ribozymes are catalytically active RNA molecules capable of cleaving RNA molecules and specific target sequences. By means of recombinant DNA techniques it is possible to alter the specificity of ribozymes. There are various classes of ribozymes. For practical applications aiming at the specific cleavage of the transcript of a certain gene, use is preferably made of representatives of two different groups of ribozymes. The first group is made up of ribozymes which belong to the group I intron ribozyme type. The second group consists of ribozymes which as a characteristic structural feature exhibit the so-called "hammerhead" motif. The specific recognition of the target RNA molecule may be modified by altering the sequences flanking this motif. By base pairing with sequences in the target molecule these sequences determine the position at which the catalytic reaction and therefore the cleavage of the target molecule takes place. Since the sequence requirements for an efficient cleavage are low, it is in principle possible to develop specific ribozymes for practically each desired RNA molecule.

In order to produce DNA molecules encoding a ribozyme which specifically cleaves transcripts of a gene encoding a Raf protein, for example a DNA sequence encoding a

catalytic domain of a ribozyme is bilaterally linked with DNA sequences which are homologous to sequences encoding the target protein. Sequences encoding the catalytic domain may for example be the catalytic domain of the satellite DNA of the SCMo virus (Davies, Virology 177 (1990), 216-224 and Steinecke, EMBO J. 11 (1992), 1525-1530) or that of the satellite DNA of the TobR virus (Haseloff and Gerlach, Nature 334 (1988), 585-591). The DNA sequences flanking the catalytic domain are preferably derived from the above-described polynucleotides contained in a pharmaceutical composition of the invention. The expression of ribozymes in order to decrease the activity in certain proteins in cells is also known to the person skilled in the art and is, for example, described in EP-B1 0 321 201.

Further potential antagonist compounds to Raf proteins include small molecules which bind to and occupy the active site of the protein thereby making the catalytic site inaccessible to substrate such that normal biological activity is prevented. Examples of small molecules include small peptides or peptide-like molecules.

Preferably, said compounds having an antagonistic effect on the polypeptide having Raf activity can be identified by a method for screening such compounds comprising the steps of

- (a) measuring the activity of said polypeptide in the presence and in the absence of the compound to be tested; and
- (b) identifying a compound acting as an antagonist if the activity measured in step (a) is lower in the presence of the compound than in the absence of the compound.

This method can be carried out as described above with regard to screening of agonist compounds.

The pharmaceutical composition of the present invention may further comprise a pharmaceutically acceptable carrier and/or diluent. Examples of suitable pharmaceutical carriers are well known in the art and include phosphate buffered saline solutions, water, emulsions, such as oil/water emulsions, various types of wetting agents, sterile solutions etc. Compositions comprising such carriers can be formulated by well known conventional methods. These pharmaceutical compositions

can be administered to the subject at a suitable dose. Administration of the suitable compositions may be effected by different ways, e.g., by intravenous, intraperitoneal, subcutaneous, intramuscular, topical, intradermal, intranasal or intrabronchial administration. The dosage regimen will be determined by the attending physician and clinical factors. As is well known in the medical arts, dosages for any one patient depends upon many factors, including the patient's size, body surface area, age, the particular compound to be administered, sex, time and route of administration, general health, and other drugs being administered concurrently. A typical dose can be, for example, in the range of 0.001 to 1000 μg (or of nucleic acid for expression or for inhibition of expression in this range); however, doses below or above this exemplary range are envisioned, especially considering the aforementioned factors. Generally, the regimen as a regular administration of the pharmaceutical composition should be in the range of 1 μg to 10 mg units per day. If the regimen is a continuous infusion, it should also be in the range of 1 μg to 10 mg units per kilogram of body weight per minute, respectively. Progress can be monitored by periodic assessment. Dosages will vary but a preferred dosage for intravenous administration of DNA is from approximately 10^6 to 10^{12} copies of the DNA molecule. The compositions of the invention may be administered locally or systemically. Administration will generally be parenterally, e.g., intravenously; DNA may also be administered directly to the target site, e.g., by biolistic delivery to an internal or external target site or by catheter to a site in an artery. Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral vehicles include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's, or fixed oils. Intravenous vehicles include fluid and nutrient replenishers, electrolyte replenishers (such as those based on Ringer's dextrose), and the like. Preservatives and other additives may also be present such as, for example, antimicrobials, anti-oxidants, chelating agents, and inert gases and the like. Furthermore, the pharmaceutical composition of the invention may comprise further agents such as interleukins or interferons depending on the intended use of the pharmaceutical composition.

In a further aspect, the present invention relates to the use of any of the compounds as described above for the preparation of a pharmaceutical composition for preventing or treating a pathological condition in which endothelial cells are involved or affected.

The term "endothelial cells" denotes cells which line and form the inner wall of all blood and lymphatic vessels. Endothelial progenitor cells on the other hand can be inside tissues, e.g. in the bone marrow or in circulation, and can be defined as endothelial cells by their ability to incorporate in blood vessels.

The term "pathological condition in which endothelial cells are involved or affected" refers in this context to any acute or chronic pathological condition, wherein endothelial cells are altered compared to the healthy state, said alteration may include cytological features, such as subcellular structure, cell morphology, the expression level(s) of one or more genes, physiological activity, molecular features such as the presence or absence of cellular markers at the cellular surface and/or proliferative activity.

Preferably this use refers to pathological conditions which are accompanied by an altered gene expression in endothelial cells compared to endothelial cells of a healthy subject. More preferred are pathological conditions wherein such an altered gene expression concerns genes the expression of which is regulated by cAMP and/or retinoic acid (RA), in particular on the transcriptional level which can, for example, be measured by determining the amount of specific RNA in the cells. Examples for cAMP and RA-dependent genes are given in Hatzopoulos (Development 125 (1998), 1457-1468) or in the Examples.

In a preferred embodiment, the above mentioned use refers to pathological conditions that can be treated by inhibiting or promoting angiogenesis.

As outlined above, the present invention is based on the observation that in B-Raf KO cells as well as in B-Raf deficient mouse embryos the expression of a lot of genes is altered compared to wild-type cells or embryos. One such alteration concerns genes which are induced in wild-type endothelial cells during angiogenesis such as Flk-1 which encodes the receptor for the vascular endothelial growth factor (VEGF) and the serine/threonine kinase receptor Alk-1. Expression of these genes is

significantly down-regulated in Raf KO cells and embryos. Consequently, modulators of B-Raf activity may be useful for inhibiting or promoting angiogenesis. Specifically, those of the above described compounds that are suited for activating B-Raf, i.e. polynucleotides, vectors, host cells, polypeptides or agonists, in the following also referred to as "activating compounds", may be useful for promoting angiogenesis. On the other hand, those of the above compounds which are antagonists, i.e. being capable of inhibiting B-Raf activity, may be useful for inhibiting angiogenesis.

A further group of genes having an altered expression in B-Raf KO cells and embryos comprises genes involved in apoptosis such as bcl-2. Apoptosis-related genes may also be used for modulating, preferably for inhibiting angiogenesis. For instance, bcl-2 is an anti-apoptosis acting gene. Bcl-2 activity prevents apoptosis in mammalian cells. Bcl-2 interacts with Raf proteins and guides them to the mitochondrial membrane. Inhibition of Raf can then induce endothelial-specific apoptosis that can be used to stop angiogenesis by selectively killing activated endothelial cells. All the diseases that angiogenesis is involved in could be treated by inducing endothelial apoptosis in the angiogenic areas.

Accordingly, in a further preferred embodiment, inhibiting or promoting of angiogenesis may be useful for preventing or treating tumors, diabetic retinopathy, chronic inflammatory diseases, such as psoriasis or arthritis, atherosclerosis or for promoting wound healing or for enhancing circulation, e.g. in conditions where circulation is impaired such as for example after heart infarct, strokes or in the extremities of older people.

Preferably, the inhibitory effect on angiogenesis triggered by antagonist compounds may be used to prevent or treat tumors, treat chronic inflammatory reactions such as psoriasis and arthritis, prevent blindness in diabetic retinopathy and to stop formation and growth of atherosclerotic plaques.

Also preferred is to use the activating compounds that are capable of promoting angiogenesis for restoring blood supply to myocardium after a heart infarct, restoring blood supply to brain after stroke, enhance circulation especially in the extremities of older people and enhance wound healing following injury.

In a further preferred embodiment, the invention refers to the above mentioned uses which are for preventing or treating inflammation or atherosclerosis.

Another group of genes the transcription and expression of which is altered in B-Raf KO cells and embryos are genes involved in endothelial-endothelial cell interactions (e.g. VE-cadherin) or endothelial-immune system cell interactions (e.g. V-CAM-1, E-selectin). These proteins are involved in the interaction of endothelial cells with leukocytes and are instrumental in recruiting these cells during inflammatory conditions. The same genes are also activated in injured endothelium during atherosclerosis. In a similar fashion, it can be expected that modulating B-Raf could lead to therapeutic intervention during inflammation and atherosclerosis.

An additional preferred embodiment relates to the uses mentioned above wherein said pathological condition can be treated by modulating the permeability of the blood-brain barrier.

Genes that control endothelial cell-cell adhesion such as VE-cadherin, E-cadherin and possibly the E-cadherin repressor factor Snail, are important for maintaining the blood-brain barrier. Therefore, modulation of Raf activity could either weaken endothelial cell adhesion and potentially circumvent the barrier for pharmacological administration of brain drugs, or strengthen the barrier to prevent toxic compound penetration to the brain tissue.

Another preferred embodiment refers to uses wherein the pathological condition can be treated by blocking or enhancing endothelial cell migration during angiogenesis or tissue remodeling.

The term "tissue remodeling" used herein refers to growth, differentiation, migration and reorganization of connective and epidermal tissue (fibroblasts, keratinocytes), blood cells (macrophages, lymphocytes, mast cells) and endothelial cells in order to heal an injured area.

The present preferred embodiment is based on the finding that another group of genes having an altered gene expression in B-Raf KO cells and embryos comprise genes involved in endothelial cell-extracellular matrix interactions, such as the gene encoding laminin gamma1, Tie-1 and TGF beta receptor II. Modulators of B-Raf could therefore block or enhance endothelial cell migration or wound healing.

Moreover, based on the altered gene expression of BMP1 and TIMP2 in B-Raf KO embryos and null cells, modulators of B-Raf can be used for inhibiting tumor development, in particular by stopping tumor cell invasion during metastasis.

Furthermore, another preferred embodiment relates to the above uses which are for treating pathophysiological conditions or injury of the vascular wall.

The term "vascular wall" refers to the inner cellular surface of blood and lymphatic vessels.

The term "pathophysiological conditions or injury" of the vascular wall refers to formation of atherosclerotic plaques and may for example include diseases such as atherosclerosis, high cholesterol levels, abnormal lipid metabolism or smoking addiction and to vascular injury due to stripping away of endothelial cells following catheterization, for instance during balloon angioplasty.

A further group of genes having an altered expression in B-Raf KO cells and embryos comprises genes involved in atherosclerosis and oxidative stress such as ApoE, plasma glutathione peroxidase, glutathione S transferases, PDGF receptors alpha and beta, and eNOS. Modulators of B-Raf could have a beneficial effect in protecting the vascular wall during pathophysiological conditions and injury. Besides the endothelial cells, such molecules could be used for treating diseases such as Alzheimer's where abnormal ApoE expression is one cause for the disease.

In a further aspect the invention relates to uses which are for preventing coagulation or fibrin deposition in the vessels.

This embodiment is based on the observation that a further group having an altered expression in B-Raf KO cells and embryos comprise genes involved in thrombosis and coagulation such as von Willebrand Factor (vWF), thrombomodulin etc. Modulators of B-Raf could thus have a beneficial effect in preventing coagulation and fibrin deposition.

Accordingly, in a further preferred embodiment said uses refer to preventing or treating stroke or myocardial infarction.

Additionally, a further preferred embodiment relates to uses which are for preventing or treating restenosis.

Further genes that were revealed to have an altered gene expression in B-Raf KO cells and embryos refer to genes involved in new functions in endothelial cells that are not sufficiently understood at the moment. For example, the expression of a number of wnt genes, their receptors and downstream wnt signaling molecules is deregulated. Although, no wnt gene functions are known in endothelial cells, a recent publication (Dennis, J. Cell Sci. 112 (1994), 3815-3820; Mao, Arterioscler. Thromb. Vasc. Biol. 20 (2000), 43-51) showed that some of the wnt receptors are induced in endothelial and smooth muscle cells during injury of endothelial cells following balloon angioplasty of occluded blood vessels. Since the major problem of this treatment is the occurring restenosis of the affected blood vessels, B-Raf modulators could prevent restenosis.

In a further embodiment, the present invention relates to the above uses, which are for preventing or treating cardiomyopathy.

A further group of genes having an altered expression in B-Raf KO cells and embryos comprises genes encoding proteins involved in heart development such as GATA-4 and 6. These two proteins are essential for heart development and are involved in myocardium differentiation. GATA-4 and 6 inhibitors or activators might be used to treat cardiomyopathies since they are known to regulate transcription of genes such as cardiac myosin heavy chain, troponin I etc. (Charron and Nemer, Cell and Dev. Biol. 10 (1999), 85-91).

In a further aspect, the present invention refers to a method for screening compounds to identify those which act as agonists or antagonists of the polypeptide as defined above, comprising the steps of

- (a) measuring the activity of said polypeptide in the presence and in the absence of the compound to be tested; and
- (b) determining that the activity measured in step (a) is
 - (i) higher in the presence of the compound than in the absence of the compound, thereby identifying a compound acting as an agonist;
 - or

- (ii) lower in the presence of the compound than in the absence of the compound, thereby identifying a compound acting as an antagonist.

The conditions under which such a screening method can be conducted have already been described above.

Furthermore, the invention relates to a method for screening compounds to identify those which act as agonists or antagonists of a polypeptide as defined in above, comprising the steps of

- (a) incubating cells which are transfected with a polynucleotide as defined above and express said polynucleotide in the presence and in the absence of a compound to be tested;
- (b) measuring a specific gene expression in the cells of (a); and
- (c) determining that the specific gene expression measured in step (b) is
 - (i) higher in the presence of the compound than in the absence of the compound, thereby identifying a compound acting as an agonist; or
 - (ii) lower in the presence of the compound than in the absence of the compound, thereby identifying a compound acting as an antagonist.

The term "specific gene expression" refers to the expression of the genes identified in Table 1. For screens of small numbers of pre-selected compounds, RNA will be isolated and analyzed by RT-PCR as described in the methods below. For high throughput screens, cells may be employed where a gene of interest has been replaced by homologous recombination with an easily quantifiable marker such as bacterial beta-galactosidase. For example, in connection with the invention, cells have been produced where one Flk-1 allele is replaced by beta-galactosidase (knock-in strategy). When Flk-1 is induced or repressed by Raf modulator, the levels of bacterial beta-galactosidase will change correspondingly. Alternatively and for gene targets where no knock-in is available, constructs can be engineered with the beta-galactosidase gene under the control of relevant gene promoter elements and embryonic endothelial cells transfected therewith.

A further aspect of the present invention relates to a method for the production of a pharmaceutical composition comprising the steps of one of the above-described

methods for screening compounds and furthermore the step of formulating the identified compound in a pharmaceutically acceptable form.

The step of formulating a compound in a pharmaceutical acceptable form so as to obtain a pharmaceutical composition of the present invention have already been described in detail above.

Additionally, the present invention relates to a diagnostic composition comprising a polynucleotide as defined above, a host cell genetically engineered with said polynucleotide or a vector comprising said polynucleotide, a polypeptide encoded by said polynucleotide or an antibody specifically recognizing said polypeptide.

The diagnostic composition optionally comprises suitable means for detection. The (poly)peptides and antibodies described above are, for example, suited for use in immunoassays in which they can be utilized in liquid phase or bound to a solid phase carrier. Examples of immunoassays which can utilize said (poly)peptide are competitive and non-competitive immunoassays in either a direct or indirect format. Examples of such immunoassays are the radioimmunoassay (RIA), the sandwich (immunometric assay) and the Western blot assay. The (poly)peptides and antibodies can be bound to many different carriers and used to isolate cells specifically bound to said polypeptides. Examples of well-known carriers include glass, polystyrene, polyvinyl chloride, polypropylene, polyethylene, polycarbonate, dextran, nylon, amyloses, natural and modified celluloses, polyacrylamides, agaroses, and magnetite. The nature of the carrier can be either soluble or insoluble for the purposes of the invention.

There are many different labels and methods of labeling known to those of ordinary skill in the art. Examples of the types of labels which can be used in the present invention include enzymes, radioisotopes, colloidal metals, fluorescent compounds, chemiluminescent compounds, and bioluminescent compounds; see also the embodiments discussed hereinabove.

Said diagnostic compositions may also be used for methods for detecting expression of a gene encoding a Raf protein by detecting the presence of mRNA coding for a membrane bound fusion protein which comprises obtaining mRNA from a cell and contacting the mRNA so obtained with a probe comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a nucleic acid molecule

as described above under suitable hybridizing conditions (see also supra), detecting the presence of mRNA hybridized to the probe, and thereby detecting the expression of the fusion protein by the cell.

Furthermore, the host cells described above may act as biosensors of early disease signs. When transplanted into a subject, such cells may monitor pathologically high or low Raf activity. Methods to construct corresponding transfected cells which allow for in vivo monitoring are well known to the person skilled in the art.

The components of the diagnostic composition of the present invention may be packaged in containers such as vials, optionally in buffers and/or solutions. If appropriate, one or more of said components may be packaged in one and the same container. Additionally or alternatively, one or more of said components may be adsorbed to a solid support such as, e.g., a nitrocellulose filter or nylon membrane, or to the well of a microtitre-plate.

Furthermore, the present invention refers in another aspect to a method for diagnosing a pathological condition or a susceptibility to a pathological condition in a subject comprising the steps of

- (a) determining the expression level of a polypeptide as defined above in a sample comprising endothelial cells or a lysate thereof; and
- (b) diagnosing a pathological condition or a susceptibility to a pathological condition if the expression level determined in step (a) is altered compared to a standard expression level.

Samples for carrying out this method may be taken according to routine clinical procedures known in the art. For instance, during by-pass operations leg veins are removed and are used for heart by-pass operations. Portions of these veins can be used for endothelial isolation. Peripheral arteries can also be removed and used, e.g., for endothelial RNA preparation. In addition, tumor specimens contain activated endothelium. Most frequently though, umbilical veins are used for isolating and growing endothelial cells.

These and other embodiments are disclosed and obvious to a skilled person and embraced by the description and the examples of the present invention. Additional literature regarding one of the above-mentioned methods, means and applications,

which can be used within the meaning of the present invention, can be obtained from the state of the art, for instance from public libraries for instance by the use of electronic means. This purpose can be served inter alia by public databases, such as the "medline", which are accessible via internet, for instance under the address <http://www.ncbi.nlm.nih.gov/PubMed/medline.html>. Other databases and addresses are known to a skilled person and can be obtained from the internet, for instance under the address <http://www.lycos.com>. An overview of sources and information regarding patents and patent applications in biotechnology is contained in Berks, TIBTECH 12 (1994), 352-364.

All of the above cited disclosures of patents, publications and database entries are specifically incorporated herein by reference in their entirety to the same extent as if each such individual patent, publication or entry were specifically and individually indicated to be incorporated by reference.

Figure 1 shows examples of genes that are deregulated in B-Raf null cells. The amount of specific mRNA has been determined in wild-type endothelial cells ("wild-type") and B-Raf null cells ("KO") using RT-PCR. For each gene, gene expression was measured with (+) and without (-) prior addition of cAMP and retinoic acid (RA) according to the protocol outlined in Hatzopoulos (Development 125 (1998), 1457-1468). As a conclusion from the results, the expression of the genes shown, in particular the cAMP and RA-mediated gene expression, is impaired in B-Raf deficient endothelial cells.

Figure 2 shows that cAMP-regulated genes are downregulated in B-Raf KO embryos whereas genes that are not regulated by cAMP and whose expression is not affected in B-Raf KO cells is also not altered in B-Raf KO embryos. The match between in vitro and in vivo data suggests that B-Raf is a main regulator of these genes in endothelial cells.

Figure 3 shows that cAMP-induced activation of the MAPK pathway in endothelial cells requires B-Raf. For detailed explanations see Example 3.

Figure 4 shows electron micrographs from mouse embryos at day 12.5. These cross-sections show that B-Raf is required for the proper development of peri-endothelial structures. Further explanations are given in Example 4. The dots in the arrows in the two upper panels mark the areas of the smooth muscle cells. The arrows point to the endothelial cells. The arrows in the bottom panels point to collagen fibers in the peri-vascular areas.

Table 1 shows target genes of the B-Raf signaling pathway

The following Examples illustrate the invention.

Experimental setup

Isolation of B-Raf null angioblasts

Mouse embryonic endothelial progenitor cells or angioblasts were isolated as previously described (Hatzopoulos, Development 125 (1988), 1457-1468). To this end, a timed pregnancies from B-Raf heterozygote mice was set up and embryos were isolated at day 7.5 for tissue culture. The mice are of a Black C57 background that are bred in our own animal facility. The original breeding pairs were kindly provided by Dr. Andreas Zimmer from NIH, Bethesda, Maryland. Each embryo was dissociated with trypsin and cultured separately on a feeder layer of mouse embryonic fibroblasts using ESCM 20% as medium (Hatzopoulos, Development 125 (1988), 1457-1468). After several months, six independent angioblast cell lines were established, each derived from a single embryo. Genomic DNA was isolated from all lines by lysing cells O/N at 55°C in 400µl 50mM KCl, 1.5mM MgCl₂, 0.45% NP40, 0.45% Tween 20 and 100µg/ml proteinase K. The lysates were heat-inactivated at 95°C for 20 minutes, and spun at 13,000 rpm in a table microfuge. The clear supernatant was collected and used for genotyping. Genotyping was performed by PCR using primer pairs that can distinguish between the wild type and the knocked out allele. Following oligonucleotides were used for genotyping:

1. mBraf3-1

5' - GCC TAT GAA GAG TAC ACC AGC AAG CTA GAT GCC C - 3' (SEQ ID NO: 7);

2. mrafB-des

5' - TAG GTT TCT GTG GTG ACT TGG GGT TGT TCC GTG A - 3' (SEQ ID NO: 8); and

3. Neo-L

5' - AGT GCC AGC GGG GCT GCT AAA - 3' (SEQ ID NO: 9).

Primers 1 and 2 amplify the wt allele and primers 1 and 3 amplify the KO allele. The PCR contained dNTPs each at 0.22mM, 1X buffer B (PROMEGA), gene-specific primers 0.5µM each, 2µl of genomic DNA and 0.6 units Taq polymerase. The PCR program was 1 minute 95°C, 1 minute 58°C, 2 minutes 72°C for 30 cycles. The reactions were given a final incubation of 10 minutes at 72°C and cooled to 4°C. The PCR products were separated on 1.5% agarose gels, the gels were stained with EtBr and photographed.

The results showed that one line lacked both wild type (wt) B-Raf alleles and it was therefore a null B-Raf endothelial cell line. The B-Raf null cells were frozen in liquid nitrogen.

Example 1: Gene expression profile analysis of wt and B-Raf null endothelial cells

In order to investigate the function of B-Raf in endothelial cells, gene expression profiles between the two different cell types were analyzed using commercially available cDNA arrays (CLONTECH) following the manufacturer's specifications. A number of the observed differences were selected for further confirmation by RNA analysis using RT-PCR. To this end, RNA was isolated from wt and B-Raf null cells with the QIAGEN RNeasy kit. 1µg of total RNA was annealed to 100ng of oligo(dT)₁₅ at 65°C for 5 minutes. Then, dNTPs (final concentration 1mM each), reaction buffer (final concentration 60mM KCl, 15mM Tris-Cl pH 8.4, 3mM MgCl₂, 0.3% Tween 20, 5mM DTT, 5mM β-MSH, 10 units RNasin) and 100 units Mo-MLV Reverse Transcriptase (LifeTechnologies) was added. First strand synthesis was then

performed at 37°C for 55 minutes. The reaction was stopped by heat inactivation at 95°C for 5 minutes.

The prepared cDNA was used as template in PCR reactions in order to analyze and compare gene expression profiles in wt and B-Raf null cells. The PCR conditions were as follows: dNTPs each at 0.22mM, 5% DMSO, 1X buffer B (PROMEGA) 50.6mM KCl, 1.53mM MgCl₂, gene-specific primers 0.5μM each, 10ng cDNA and 0.6 units Taq polymerase). The PCR program was 1 minute 95°C, 1 minute 65°C, 1 minute 72°C for 30 cycles. After cycling, the reactions were incubated for 5 minutes at 72°C and cooled to 4°C. Amplification products were separated on 1.5% agarose gels, the gels were stained with EtBr and photographed.

The results showed that a number of genes are deregulated, i.e. having an altered gene expression, in endothelial cells following gene inactivation of B-Raf (Figure 1). It was also discovered that genes that are normally induced by cAMP and retinoic acid (RA) in wt angioblasts (Hatzopoulos, Development 125 (1998), 1457-1468) fail to do so in the B-Raf null cells (Figure 1). These results are consistent with the role of the B-Raf protein kinase as a downstream molecule in the cAMP-signaling pathway. B-Raf is a direct target of Protein Kinase A (PKA) which in turn is activated by cAMP. Consequently, the cAMP-signaling pathway is impaired in the B-Raf null cells. Table 1 provides a list of some of the genes that were shown to have an up or down-regulated gene expression in B-Raf null cells.

Table 1

GATA-4

GATA-6

VWF

Flk-1

Alk-1

ENOS

Thrombomodulin

V-CAM1

VE-cadherin

E-Selectin

Tie-1

PDGF receptor alpha

PDGF receptor beta

Laminin gamma 1

Wnt (wnt11)

Wnt signaling pathway genes (disheveled, groucho, frizzled receptors)

TGF beta receptor II

Bcl2

Snail

E-cadherin

Plasma glutathione peroxidase (GPX)

Glutathione S transferase (GST)

ApoE

BMP1

TIMP2

Example 2: Endothelial gene expression repertoire is altered in B-Raf deficient mice

In subsequent experiments, the results obtained in B-Raf null cells in tissue culture were confirmed by comparing gene expression profiles between wt and B-Raf KO embryos in vivo. To this end, timed pregnancies from B-Raf heterozygote mice and isolated embryos were set up at day 10.5. A part from each embryo was used for genomic DNA preparation and genotyping as described above. The remaining tissue from each embryo was processed for RNA isolation with the RNeasy QIAGEN kit. RNA from wt and B-Raf KO embryos was then used for RT-PCR analysis as described above.

The thus obtained results showed that genes, whose expression changed in the B-Raf null cells, were similarly affected during embryonic vascular development. In brief, genes that were down regulated in null cells in tissue culture, and/or failed to be induced by cAMP and retinoic acid (RA), were also markedly reduced in the B-Raf deficient embryos (Figure 2).

Example 3: B-Raf kinase cascade assay with wt and B-Raf null embryonic endothelial progenitor cells (eEPC)

Wild type or B-Raf KO eEPCs were grown in 10 cm tissue culture plates to a confluence of 80 % and starved in 0.1 % BSA-DMEM for 18 h. The cells were subsequently stimulated for 5 or 10 minutes with 0.1 % BSA-DMEM containing 0.5mM cAMP. In control reactions the 0.5mM cAMP was omitted. Following stimulation, the cells were lysed in 800 µl Lysis buffer per plate.

(Lysis buffer: 20 mM Tris pH 7.5

5 mM MgCl₂

10 mM EGTA

150 mM NaCl

50 mM NaF

40 mM Na₂P₂O₇

1 mM Na₃VO₄

1 % Triton

1 mM PMSF, Aprotinin, Leupeptin)

The cells were incubated for 10 min on ice and the lysates were centrifuged at 12,000 rpm, 15 min at 4°C and protein concentration was determined in the supernatant using the BIORAD protein assay kit. 500µg of protein lysate were pre-cleared with addition of 1/10 Volume of Protein-G-Sepharose and incubated for 10 min at 4°C. The samples were centrifuged at 12,000 rpm, 10 min, 4°C. The B-Raf protein was immunoprecipitated (IP) as follows: 2 µg/ml of Anti-B-Raf antibody (from St. Cruz Biotechnology, St. Cruz, CA, USA) were added and the lysates were incubated for 2 hours at 4°C. Subsequently, 50µl of 50% beads-slurry of Protein-G-Sepharose (in Lysis buffer or PBS) were added and incubated for another hour. The beads were washed 3x in lysis buffer without protease inhibitors (PMSF, Aprotinin, Leupeptin) at 4°C and resuspended in immunoprecipitation buffer (1% NP-40, 0.5% sodium dioxcholate, 0.1% SDS in PBS).

B-Raf activity was measured using the B-Raf kinase kit from Upstate Biotechnologies: ADB and Mg/ATP-Mix were thawed rapidly, mixed thoroughly and placed on ice. A Master-Mix was prepared on ice as follows:

- 10 µl Mg/ATP-Mix
- 1.6 µl MEK1, inactive
- 4 µl MAP Kinase 2/Erk2
- 2 µl [γ -³²P]-ATP-solution
- 22.4 µl ADB (total volume per reaction 40 µl)

40 µl of Master-Mix was dispensed to each tube containing the B-Raf immunoprecipitate (on ice), mixed well, and the tubes were transferred to a shaker at 30°C. As positive control 1 µl of activated B-Raf protein (provided with the kit) was used. The reactions were incubated for 30 min and terminated by transferring on ice and subsequent addition of Laemmli loading buffer. The samples were boiled for 5 min at 95°C and analysed on a standard polyacrylamide gel. The gel was blotted on a PVDF-Membrane and B-Raf activity was detected by autoradiography and visualization of phosphorylated MEK1 and MAPK2 (ERK2) (Figure 3).

As it is shown in Figure 3, cAMP can induce activation of the MAP kinase pathway in eEPCs as it is apparent from the phosphorylation of MEK1 and MAPK2 (also called ERK2). This activation fails to occur in eEPCs lacking B-Raf. These results suggest that the effects of a number of divers extracellular signals that modulate intracellular

cAMP levels in endothelial cells, and thus lead to MAPK pathway activation, can in turn be modulated by agents that affect B-Raf activity.

Example 4: Electron microscopy analysis of wild type and B-Raf KO embryos

A pair of B-Raf heterozygote mice was mated and embryos were isolated at day 12.5. The extra-embryonic yolk sac membrane was placed in DNA lysis buffer and genotyped for identification of wild type, heterozygote and homozygote embryos (the genotyping protocol has been described above in connection with the isolation of B-Raf null angioblasts). The corresponding embryos were individually fixed immediately upon isolation in 2-4% ice-cold paraformaldehyde in PBS O/N. Following genotyping and identification, wild type and homozygote embryos were paraffin embedded and sectioned. Sections were then analyzed using electron microscopy (Figure 4).

Electron microscopy of wild type and KO embryos revealed that B-Raf is important for the proper establishment of the endothelial / peri-endothelial environment. Blood vessels in B-Raf KO embryos, such as the dorsal aorta, display a highly disorganized smooth muscle around the inner endothelial cell layer (Figure 4). It was also evident that the KO tissues had dramatically reduced collagen fibers around the blood vessels. This phenotype is strikingly similar to the phenotype of the angiopoietin 1 (Ang1) KO mice (Suri, Cell 87 (1996), 1171-1180). Ang1 is a ligand of the endothelial-specific tyrosine kinase receptor tie-2. Tie-2 is critical for endothelial cell interactions with peri-vascular cells such as smooth muscle cells and it is involved in angiogenesis, wound healing and vascular remodeling (Dumont, Gen. Develop. 8 (1994), 1897-1909; Suri, Science 282 (1998), 468-471; Papapetropoulos, J. Biol. Chem. 275 (2000), 9102-9105). The findings presented herein indicate that B-Raf is downstream from tie-2 and thus further substantiate the role of B-Raf in angiogenesis, wound healing and endothelial cell migration.

CLAIMS

1. A pharmaceutical composition comprising a compound selected from the group consisting of
 - (i) polynucleotides encoding a Raf protein selected from the group consisting of
 - (a) polynucleotides encoding a polypeptide comprising the amino acid sequence of any one of SEQ ID NOs: 2, 4 and 6;
 - (b) polynucleotides comprising the coding region of the nucleotide sequence of any one of SEQ ID NOs: 1, 3 and 5;
 - (c) polynucleotides encoding a fragment of a polypeptide encoded by a polynucleotide of (a) or (b); and
 - (d) polynucleotides the complementary strand of which hybridizes with a polynucleotide of any one of (a) to (c) and encode a polypeptide having Raf activity;
 - (ii) vectors comprising and capable of expressing said polynucleotide;
 - (iii) host cells genetically engineered with said polynucleotide or said vector;
 - (iv) polypeptides encoded by said polynucleotide; and
 - (v) agonists of a polypeptide encoded by said polynucleotide;and optionally a pharmaceutically acceptable carrier.
2. The pharmaceutical composition of claim 1, wherein the Raf protein is B-Raf.
3. A pharmaceutical composition comprising a compound which is an antagonist of the polypeptide encoded by a polynucleotide as defined in claim 1 or 2 and optionally a pharmaceutically acceptable carrier.
4. The pharmaceutical composition of claim 1 or 2, wherein the compound which is an agonist can be identified by a method for screening such compounds comprising the steps of
 - (a) measuring the activity of said polypeptide in the presence and in the absence of the compound to be tested; and

- (b) identifying a compound acting as an agonist if the activity measured in step (a) is higher in the presence of the compound than in the absence of the compound.
- 5. The pharmaceutical composition of claim 3, wherein the compound which is an antagonist can be identified by a method for screening such compounds comprising the steps of
 - (a) measuring the activity of said polypeptide in the presence and in the absence of the compound to be tested; and
 - (b) identifying a compound acting as an antagonist if the activity measured in step (a) is lower in the presence of the compound than in the absence of the compound.
- 6. Use of a compound as defined in any one of claims 1 to 5 for the preparation of a pharmaceutical composition for preventing or treating a pathological condition in which endothelial cells are involved or affected.
- 7. The use of claim 6, wherein the pathological condition can be treated by inhibiting or promoting angiogenesis.
- 8. The use of claim 6 or 7, which is for preventing or treating tumors, diabetic retinopathy or chronic inflammatory disease or for promoting wound healing or for enhancing circulation.
- 9. The use of claim 6, which is for preventing or treating inflammation or atherosclerosis.
- 10. The use of claim 6, wherein the pathological condition can be treated by modulating the permeability of the blood-brain barrier.
- 11. The use of claim 6, wherein the pathological condition can be treated by blocking or enhancing endothelial cell migration during angiogenesis or tissue remodeling.

12. The use of claim 6, which is for inhibiting tumor development.
13. The use of claim 12, wherein said tumor development is metastasis.
14. The use of claim 6, which is for treating pathophysiological conditions or injury of the vascular wall.
15. The use of claim 6, which is for treating Alzheimer's disease.
16. The use of claim 6, which is for preventing coagulation or fibrin deposition in the vessels.
17. The use of claim 6 or 16, which is for preventing or treating stroke or myocardial infarction.
18. The use of claim 6, which is for preventing or treating restenosis.
19. The use of claim 6, which is for preventing or treating cardiomyopathy.
20. A method for screening compounds to identify those which act as agonists or antagonists of the polypeptide as defined in claim 1 or 2, comprising the steps of
 - (a) measuring the activity of said polypeptide in the presence and in the absence of the compound to be tested; and
 - (b) determining that the activity measured in step (a) is
 - (i) higher in the presence of the compound than in the absence of the compound, thereby identifying a compound acting as an agonist;
or
 - (ii) lower in the presence of the compound than in the absence of the compound, thereby identifying a compound acting as an antagonist.
21. A method for screening compounds to identify those which act as agonists or antagonists of a polypeptide as defined in claim 1 or 2, comprising the steps of

- (a) incubating cells which are transfected with a polynucleotide as defined in claim 1 or 2 and express said polynucleotide in the presence and in the absence of a compound to be tested;
 - (b) measuring a specific gene expression in the cells of (a); and
 - (c) determining that the specific gene expression measured in step (b) is
 - (i) higher in the presence of the compound than in the absence of the compound, thereby identifying a compound acting as an agonist; or
 - (ii) lower in the presence of the compound than in the absence of the compound, thereby identifying a compound acting as an antagonist.
22. A method for the production of a pharmaceutical composition comprising the steps of the method of claim 20 or 21 and furthermore the step of formulating the identified compound in a pharmaceutically acceptable form.
23. A diagnostic composition comprising the polynucleotide as defined in claim 1 or 2, a host cell genetically engineered with said polynucleotide or a vector comprising said polynucleotide, a polypeptide encoded by said polynucleotide or an antibody specifically recognizing said polypeptide.
24. A method for diagnosing a pathological condition or a susceptibility to a pathological condition in a subject comprising the steps of
- (a) determining the expression level of a polypeptide as defined in claim 1 or 2 in a sample comprising endothelial cells or a lysate thereof; and
 - (b) diagnosing a pathological condition or a susceptibility to a pathological condition if the expression level determined in step (a) is altered compared to a standard expression level.

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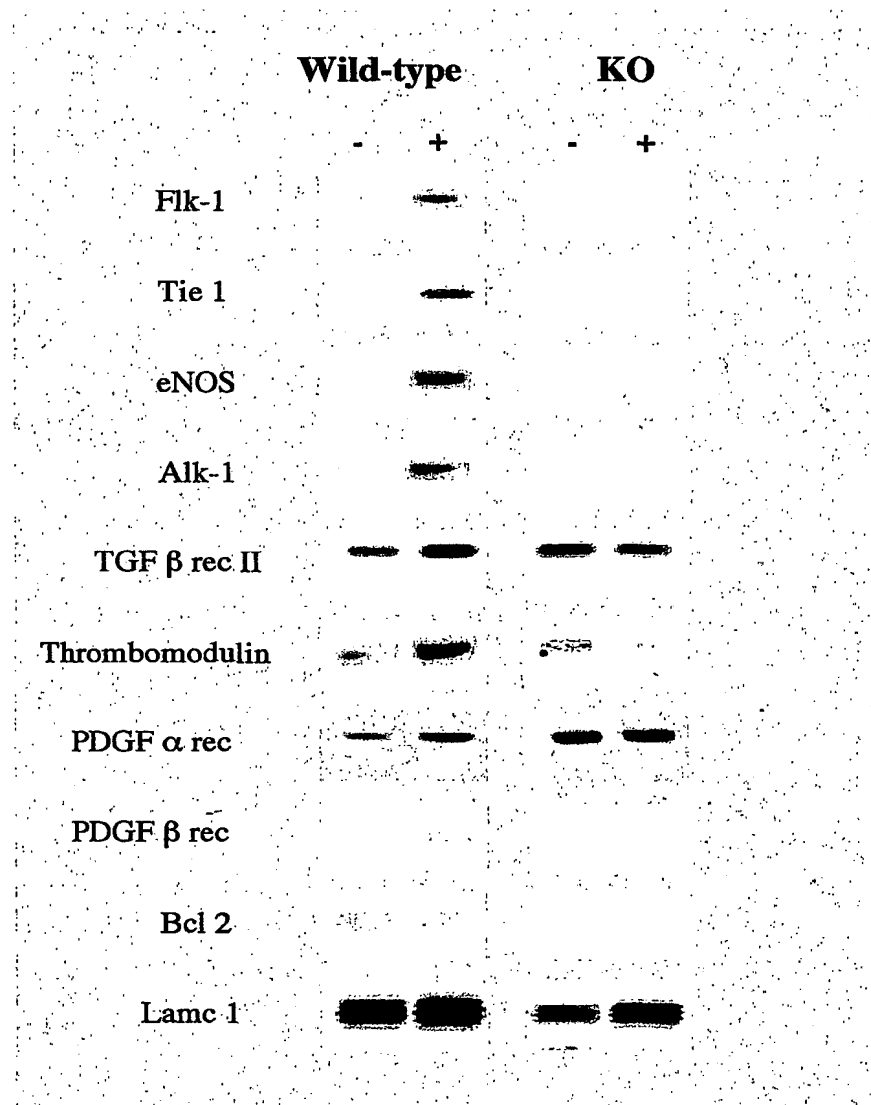


Fig. 1

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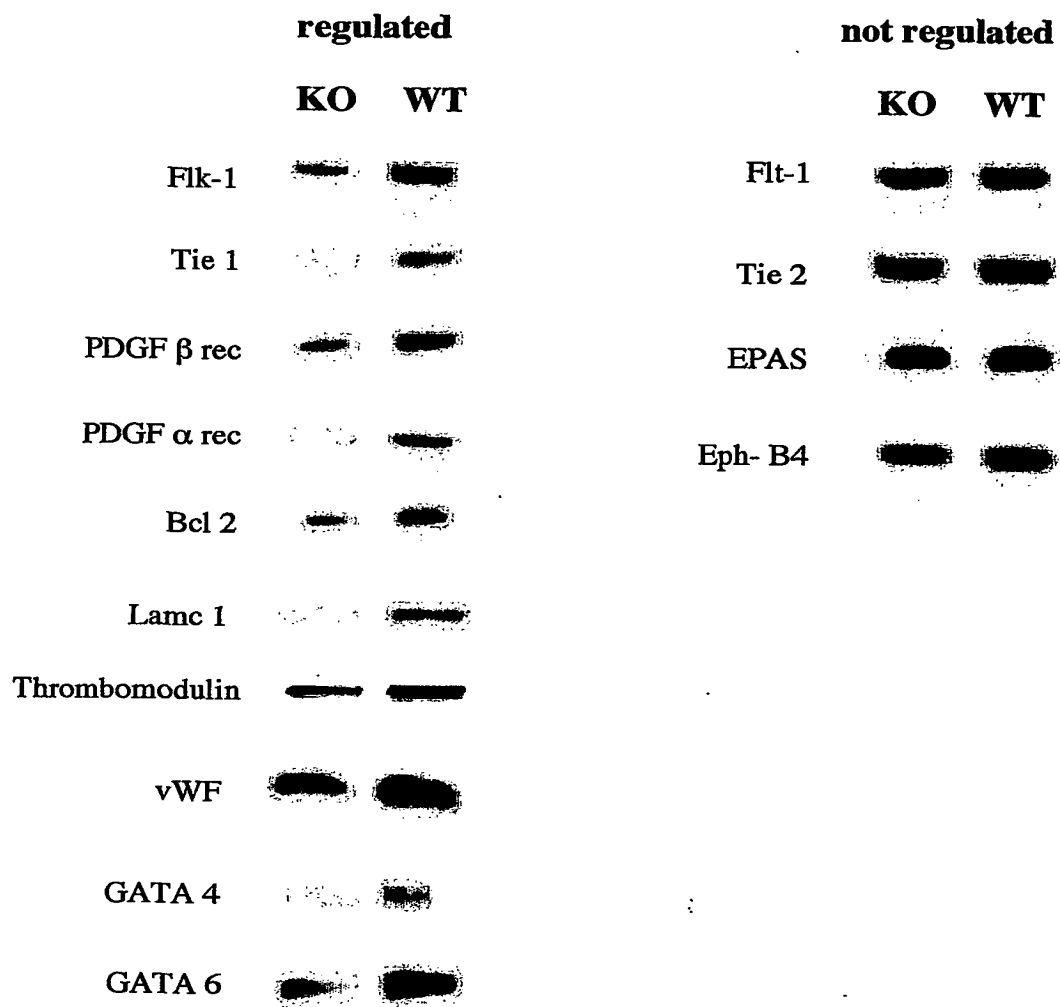


Fig. 2

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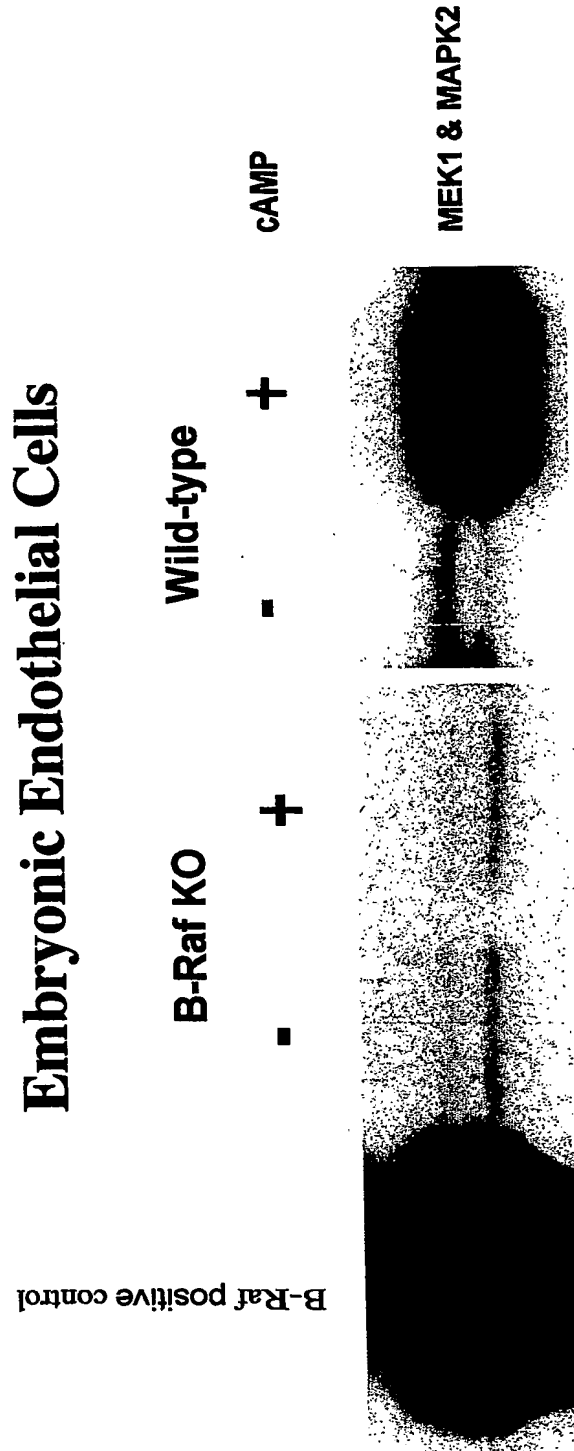


Fig. 3

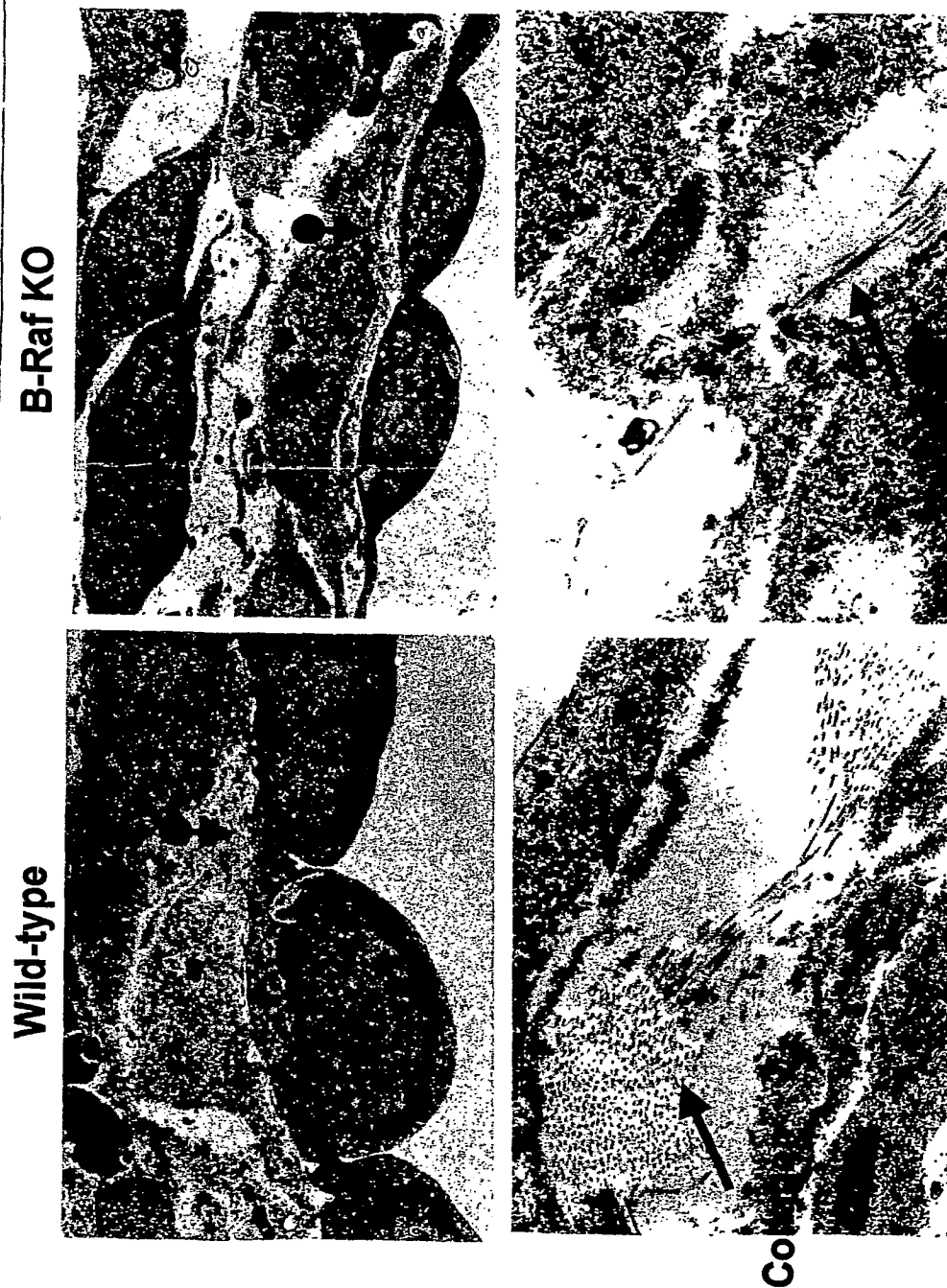


Fig. 4

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 Met Ala Ala Leu Ser Gly Gly Gly Gly Gly Gly Ala Glu Pro Gly Gln
 1 5 10 15

gct ctg ttc aac ggg gac atg gag ccc gag gcc ggc gcc ggc cgg ccc 157
 Ala Leu Phe Asn Gly Asp Met Glu Pro Glu Ala Gly Ala Gly Arg Pro
 20 25 30

gcg gcc tct tct gct gcg gac cct gcc att ccg gag gag gtg tgg aat 205
 Ala Ala Ser Ser Ala Ala Asp Pro Ala Ile Pro Glu Glu Val Trp Asn
 35 40 45

atc aaa caa atg att aag ttg aca cag gaa cat ata gag gcc cta ttg 253
 Ile Lys Gln Met Ile Lys Leu Thr Gln Glu His Ile Glu Ala Leu Leu
 50 55 60

gac aaa ttt ggt ggg gag cat aat cca cca tca ata tat ctg gag gcc 301
 Asp Lys Phe Gly Gly Glu His Asn Pro Pro Ser Ile Tyr Leu Glu Ala
 65 70 75 80

tat gaa gaa tac acc agc aag cta gat gca ctc caa caa aga gaa caa 349
 Tyr Glu Glu Tyr Thr Ser Lys Leu Asp Ala Leu Gln Gln Arg Glu Gln
 85 90 95

cag tta ttg gaa tct ctg ggg aac gga act gat ttt tct gtt tct agc 397
 Gln Leu Leu Glu Ser Leu Gly Asn Gly Thr Asp Phe Ser Val Ser Ser
 100 105 110

tct gca tca atg gat acc gtt aca tct tct tcc tct tct agc ctt tca 445
 Ser Ala Ser Met Asp Thr Val Thr Ser Ser Ser Ser Ser Ser Leu Ser
 115 120 125

gtg cta cct tca tct ctt tca gtt ttt caa aat ccc aca gat gtg gca 493
 Val Leu Pro Ser Ser Leu Ser Val Phe Gln Asn Pro Thr Asp Val Ala
 130 135 140

cgg agc aac ccc aag tca cca caa aaa cct atc gtt aga gtc ttc ctg 541
 Arg Ser Asn Pro Lys Ser Pro Gln Lys Pro Ile Val Arg Val Phe Leu
 145 150 155 160

ccc aac aaa cag agg aca gtg gta cct gca agg tgt gga gtt aca gtc 589
 Pro Asn Lys Gln Arg Thr Val Val Pro Ala Arg Cys Gly Val Thr Val
 165 170 175

cga gac agt cta aag aaa gca ctg atg atg aga ggt cta atc cca gag 637
 Arg Asp Ser Leu Lys Lys Ala Leu Met Met Arg Gly Leu Ile Pro Glu
 180 185 190

tgc tgt gct gtt tac aga att cag gat gga gag aag aaa cca att ggt 685
 Cys Cys Ala Val Tyr Arg Ile Gln Asp Gly Glu Lys Lys Pro Ile Gly
 195 200 205

tgg gac act gat att tcc tgg ctt act gga gaa gaa ttg cat gtg gaa 733
 Trp Asp Thr Asp Ile Ser Trp Leu Thr Gly Glu Glu Leu His Val Glu
 210 215 220

gtg ttg gag aat gtt cca ctt aca aca cac aac ttt gta cga aaa acg 781
 Val Leu Glu Asn Val Pro Leu Thr Thr His Asn Phe Val Arg Lys Thr
 225 230 235 240

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ttt ttc acc tta gca ttt tgt gac ttt tgt cga aag ctg ctt ttc cag	829
Phe Phe Thr Leu Ala Phe Cys Asp Phe Cys Arg Lys Leu Leu Phe Gln	
245 250 255	
ggg ttc cgc tgt caa aca tgt ggt tat aaa ttt cac cag cgt tgt agt	877
Gly Phe Arg Cys Gln Thr Cys Gly Tyr Lys Phe His Gln Arg Cys Ser	
260 265 270	
aca gaa gtt cca ctg atg tgt gtt aat tat gac caa ctt gat ttg ctg	925
Thr Glu Val Pro Leu Met Cys Val Asn Tyr Asp Gln Leu Asp Leu Leu	
275 280 285	
ttt gtc tcc aag ttc ttt gaa cac cac cca ata cca cag gaa gag gcg	973
Phe Val Ser Lys Phe Phe Glu His His Pro Ile Pro Gln Glu Glu Ala	
290 295 300	
tcc tta gca gag act gcc cta aca tct gga tca tcc cct tcc gca ccc	1021
Ser Leu Ala Glu Thr Ala Leu Thr Ser Gly Ser Ser Pro Ser Ala Pro	
305 310 315 320	
gcc tcg gac tct att ggg ccc caa att ctc acc agt ccg tct cct tca	1069
Ala Ser Asp Ser Ile Gly Pro Gln Ile Leu Thr Ser Pro Ser Pro Ser	
325 330 335	
aaa tcc att cca att cca cag ccc ttc cga cca gca gat gaa gat cat	1117
Lys Ser Ile Pro Ile Pro Gln Pro Phe Arg Pro Ala Asp Glu Asp His	
340 345 350	
cga aat caa ttt ggg caa cga gac cga tcc tca tca gct ccc aat gtg	1165
Arg Asn Gln Phe Gly Gln Arg Asp Arg Ser Ser Ser Ala Pro Asn Val	
355 360 365	
cat ata aac aca ata gaa cct gtc aat att gat gac ttg att aga gac	1213
His Ile Asn Thr Ile Glu Pro Val Asn Ile Asp Asp Leu Ile Arg Asp	
370 375 380	
caa gga ttt cgt ggt gat gga gga tca acc aca ggt ttg tct gct acc	1261
Gln Gly Phe Arg Gly Asp Gly Gly Ser Thr Thr Gly Leu Ser Ala Thr	
385 390 395 400	
ccc cct gcc tca tta cct ggc tca cta act aac gtg aaa gcc tta cag	1309
Pro Pro Ala Ser Leu Pro Gly Ser Leu Thr Asn Val Lys Ala Leu Gln	
405 410 415	
aaa tct cca gga cct cag cga gaa agg aag tca tct tca tcc tca gaa	1357
Lys Ser Pro Gly Pro Gln Arg Glu Arg Lys Ser Ser Ser Ser Ser Glu	
420 425 430	
gac agg aat cga atg aaa aca ctt ggt aga cgg gac tcg agt gat gat	1405
Asp Arg Asn Arg Met Lys Thr Leu Gly Arg Arg Asp Ser Ser Asp Asp	
435 440 445	
tgg gag att cct gat ggg cag att aca gtg gga caa aga att gga tct	1453
Trp Glu Ile Pro Asp Gly Gln Ile Thr Val Gly Gln Arg Ile Gly Ser	
450 455 460	
gga tca ttt gga aca gtc tac aag gga aag tgg cat ggt gat gtg gca	1501
Gly Ser Phe Gly Thr Val Tyr Lys Gly Lys Trp His Gly Asp Val Ala	
465 470 475 480	

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gtg aaa atg ttg aat gtg aca gca cct aca cct cag cag tta caa gcc	1549
Val Lys Met Leu Asn Val Thr Ala Pro Thr Pro Gln Gln Leu Gln Ala	
485 490 495	
ttc aaa aat gaa gta gga gta ctc agg aaa aca cga cat gtg aat atc	1597
Phe Lys Asn Glu Val Gly Val Leu Arg Lys Thr Arg His Val Asn Ile	
500 505 510	
cta ctc ttc atg ggc tat tcc aca aag cca caa ctg gct att gtt acc	1645
Leu Leu Phe Met Gly Tyr Ser Thr Lys Pro Gln Leu Ala Ile Val Thr	
515 520 525	
cag tgg tgt gag ggc tcc agc ttg tat cac cat ctc cat atc att gag	1693
Gln Trp Cys Glu Gly Ser Ser Leu Tyr His His Leu His Ile Ile Glu	
530 535 540	
acc aaa ttt gag atg atc aaa ctt ata gat att gca cga cag act gca	1741
Thr Lys Phe Glu Met Ile Lys Leu Ile Asp Ile Ala Arg Gln Thr Ala	
545 550 555 560	
cag ggc atg gat tac tta cac gcc aag tca atc atc cac aga gac ctc	1789
Gln Gly Met Asp Tyr Leu His Ala Lys Ser Ile Ile His Arg Asp Leu	
565 570 575	
aag agt aat aat ata ttt ctt cat gaa gac ctc aca gta aaa ata ggt	1837
Lys Ser Asn Asn Ile Phe Leu His Glu Asp Leu Thr Val Lys Ile Gly	
580 585 590	
gat ttt ggt cta gct aca gtg aaa tct cga tgg agt ggg tcc cat cag	1885
Asp Phe Gly Leu Ala Thr Val Lys Ser Arg Trp Ser Gly Ser His Gln	
595 600 605	
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Phe Glu Gln Leu Ser Gly Ser Ile Leu Trp Met Ala Pro Glu Val Ile	
610 615 620	
aga atg caa gat aaa aat cca tac agc ttt cag tca gat gta tat gca	1981
Arg Met Gln Asp Lys Asn Pro Tyr Ser Phe Gln Ser Asp Val Tyr Ala	
625 630 635 640	
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Phe Gly Ile Val Leu Tyr Glu Leu Met Thr Gly Gln Leu Pro Tyr Ser	
645 650 655	
aac atc aac aac agg gac cag ata att ttt atg gtg gga cga gga tac	2077
Asn Ile Asn Asn Arg Asp Gln Ile Ile Phe Met Val Gly Arg Gly Tyr	
660 665 670	
ctg tct cca gat ctc agt aag gta cgg agt aac tgt cca aaa gcc atg	2125
Leu Ser Pro Asp Leu Ser Lys Val Arg Ser Asn Cys Pro Lys Ala Met	
675 680 685	
aag aga tta atg gca gag tgc ctc aaa aag aaa aga gat gag aga cca	2173
Lys Arg Leu Met Ala Glu Cys Leu Lys Lys Lys Arg Asp Glu Arg Pro	
690 695 700	
ctc ttt ccc caa att ctc gcc tct att gag ctg ctg gcc cgc tca ttg	2221
Leu Phe Pro Gln Ile Leu Ala Ser Ile Glu Leu Leu Ala Arg Ser Leu	
705 710 715 720	

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cca aaa att cac cgc agt gca tca gaa ccc tcc ttg aat cgg gct ggt 2269
Pro Lys Ile His Arg Ser Ala Ser Glu Pro Ser Leu Asn Arg Ala Gly
              725              730              735

ttc caa aca gag gat ttt agt cta tat gct tgt gct tct cca aaa aca 2317
Phe Gln Thr Glu Asp Phe Ser Leu Tyr Ala Cys Ala Ser Pro Lys Thr
              740              745              750

ccc atc cag gca ggg gga tat ggt gcg ttt cct gtc cac tga 2359
Pro Ile Gln Ala Gly Gly Tyr Gly Ala Phe Pro Val His
              755              760              765

aacaaatgag tgagagagtt caggagagta gcaacaaaag gaaaataaat gaacatatgt 2419

ttgcttatat gttaaattga ataaaataact ctcttttttt ttaaggtgga aaaaaaaaaa 2479

aaaaaaaaaa aaaaaaaaaa aaaaaaaacc c 2510

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 <212> PRT
 <213> Homo sapiens

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 35          40          45
Ile Lys Gln Met Ile Lys Leu Thr Gln Glu His Ile Glu Ala Leu Leu
 50          55          60
Asp Lys Phe Gly Gly Glu His Asn Pro Pro Ser Ile Tyr Leu Glu Ala
 65          70          75          80
Tyr Glu Glu Tyr Thr Ser Lys Leu Asp Ala Leu Gln Gln Arg Glu Gln
 85          90          95
Gln Leu Leu Glu Ser Leu Gly Asn Gly Thr Asp Phe Ser Val Ser Ser
 100         105         110
Ser Ala Ser Met Asp Thr Val Thr Ser Ser Ser Ser Ser Ser Leu Ser
 115         120         125
Val Leu Pro Ser Ser Leu Ser Val Phe Gln Asn Pro Thr Asp Val Ala
 130         135         140
Arg Ser Asn Pro Lys Ser Pro Gln Lys Pro Ile Val Arg Val Phe Leu
 145         150         155         160
Pro Asn Lys Gln Arg Thr Val Val Pro Ala Arg Cys Gly Val Thr Val
 165         170         175
Arg Asp Ser Leu Lys Lys Ala Leu Met Met Arg Gly Leu Ile Pro Glu
 180         185         190
Cys Cys Ala Val Tyr Arg Ile Gln Asp Gly Glu Lys Lys Pro Ile Gly
 195         200         205
Trp Asp Thr Asp Ile Ser Trp Leu Thr Gly Glu Glu Leu His Val Glu
 210         215         220
Val Leu Glu Asn Val Pro Leu Thr Thr His Asn Phe Val Arg Lys Thr
 225         230         235         240
Phe Phe Thr Leu Ala Phe Cys Asp Phe Cys Arg Lys Leu Leu Phe Gln
 245         250         255
Gly Phe Arg Cys Gln Thr Cys Gly Tyr Lys Phe His Gln Arg Cys Ser
 260         265         270

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Ser	Leu	Ala	Glu	Thr	Ala	Leu	Thr	Ser	Gly	Ser	Ser	Pro	Ser	Ala	Pro
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Ala	Ser	Asp	Ser	Ile	Gly	Pro	Gln	Ile	Leu	Thr	Ser	Pro	Ser	Pro	Ser
				325					330						335
Lys	Ser	Ile	Pro	Ile	Pro	Gln	Pro	Phe	Arg	Pro	Ala	Asp	Glu	Asp	His
			340					345					350		
Arg	Asn	Gln	Phe	Gly	Gln	Arg	Asp	Arg	Ser	Ser	Ser	Ala	Pro	Asn	Val
		355					360					365			
His	Ile	Asn	Thr	Ile	Glu	Pro	Val	Asn	Ile	Asp	Asp	Leu	Ile	Arg	Asp
	370					375					380				
Gln	Gly	Phe	Arg	Gly	Asp	Gly	Gly	Ser	Thr	Thr	Gly	Leu	Ser	Ala	Thr
385					390					395					400
Pro	Pro	Ala	Ser	Leu	Pro	Gly	Ser	Leu	Thr	Asn	Val	Lys	Ala	Leu	Gln
				405					410					415	
Lys	Ser	Pro	Gly	Pro	Gln	Arg	Glu	Arg	Lys	Ser	Ser	Ser	Ser	Ser	Glu
			420					425					430		
Asp	Arg	Asn	Arg	Met	Lys	Thr	Leu	Gly	Arg	Arg	Asp	Ser	Ser	Asp	Asp
		435					440					445			
Trp	Glu	Ile	Pro	Asp	Gly	Gln	Ile	Thr	Val	Gly	Gln	Arg	Ile	Gly	Ser
	450					455					460				
Gly	Ser	Phe	Gly	Thr	Val	Tyr	Lys	Gly	Lys	Trp	His	Gly	Asp	Val	Ala
465					470					475					480
Val	Lys	Met	Leu	Asn	Val	Thr	Ala	Pro	Thr	Pro	Gln	Gln	Leu	Gln	Ala
				485					490					495	
Phe	Lys	Asn	Glu	Val	Gly	Val	Leu	Arg	Lys	Thr	Arg	His	Val	Asn	Ile
			500					505					510		
Leu	Leu	Phe	Met	Gly	Tyr	Ser	Thr	Lys	Pro	Gln	Leu	Ala	Ile	Val	Thr
		515					520					525			
Gln	Trp	Cys	Glu	Gly	Ser	Ser	Leu	Tyr	His	His	Leu	His	Ile	Ile	Glu
	530					535					540				
Thr	Lys	Phe	Glu	Met	Ile	Lys	Leu	Ile	Asp	Ile	Ala	Arg	Gln	Thr	Ala
545					550					555					560
Gln	Gly	Met	Asp	Tyr	Leu	His	Ala	Lys	Ser	Ile	Ile	His	Arg	Asp	Leu
				565					570					575	
Lys	Ser	Asn	Asn	Ile	Phe	Leu	His	Glu	Asp	Leu	Thr	Val	Lys	Ile	Gly
			580					585					590		
Asp	Phe	Gly	Leu	Ala	Thr	Val	Lys	Ser	Arg	Trp	Ser	Gly	Ser	His	Gln
		595					600					605			
Phe	Glu	Gln	Leu	Ser	Gly	Ser	Ile	Leu	Trp	Met	Ala	Pro	Glu	Val	Ile
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Arg	Met	Gln	Asp	Lys											

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 gctgcatca atg gag cac ata cag gga gct tgg aag acg atc agc aat ggt 171
 Met Glu His Ile Gln Gly Ala Trp Lys Thr Ile Ser Asn Gly
 1 5 10
 ttt gga ttc aaa gat gcc gtg ttt gat ggc tcc agc tgc atc tct cct 219
 Phe Gly Phe Lys Asp Ala Val Phe Asp Gly Ser Ser Cys Ile Ser Pro
 15 20 25 30
 aca ata gtt cag cag ttt ggc tat cag cgc cgg gca tca gat gat ggc 267
 Thr Ile Val Gln Gln Phe Gly Tyr Gln Arg Arg Ala Ser Asp Asp Gly
 35 40 45
 aaa ctc aca gat cct tct aag aca agc aac act atc cgt gtt ttc ttg 315
 Lys Leu Thr Asp Pro Ser Lys Thr Ser Asn Thr Ile Arg Val Phe Leu
 50 55 60
 ccg aac aag caa aga aca gtg gtc aat gtg cga aat gga atg agc ttg 363
 Pro Asn Lys Gln Arg Thr Val Val Asn Val Arg Asn Gly Met Ser Leu
 65 70 75
 cat gac tgc ctt atg aaa gca ctc aag gtg agg ggc ctg caa cca gag 411
 His Asp Cys Leu Met Lys Ala Leu Lys Val Arg Gly Leu Gln Pro Glu
 80 85 90
 tgc tgt gca gtg ttc aga ctt ctc cac gaa cac aaa ggt aaa aaa gca 459
 Cys Cys Ala Val Phe Arg Leu Leu His Glu His Lys Gly Lys Lys Ala
 95 100 105 110
 cgc tta gat tgg aat act gat gct gcg tct ttg att gga gaa gaa ctt 507
 Arg Leu Asp Trp Asn Thr Asp Ala Ala Ser Leu Ile Gly Glu Glu Leu
 115 120 125
 caa gta gat ttc ctg gat cat gtt ccc ctc aca aca cac aac ttt gct 555
 Gln Val Asp Phe Leu Asp His Val Pro Leu Thr Thr His Asn Phe Ala
 130 135 140
 cgg aag acg ttc ctg aag ctt gcc ttc tgt gac atc tgt cag aaa ttc 603
 Arg Lys Thr Phe Leu Lys Leu Ala Phe Cys Asp Ile Cys Gln Lys Phe
 145 150 155
 ctg ctc aat gga ttt cga tgt cag act tgt ggc tac aaa ttt cat gag 651

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Leu	Leu	Asn	Gly	Phe	Arg	Cys	Gln	Thr	Cys	Gly	Tyr	Lys	Phe	His	Glu		
160						165					170						
cac	tgt	agc	acc	aaa	gta	cct	act	atg	tgt	gtg	gac	tgg	agt	aac	atc	699	
His	Cys	Ser	Thr	Lys	Val	Pro	Thr	Met	Cys	Val	Asp	Trp	Ser	Asn	Ile		
175					180					185					190		
aga	caa	ctc	tta	ttg	ttt	cca	aat	tcc	act	att	ggg	gat	agt	gga	gtc	747	
Arg	Gln	Leu	Leu	Leu	Phe	Pro	Asn	Ser	Thr	Ile	Gly	Asp	Ser	Gly	Val		
				195				200						205			
cca	gca	cta	cct	tct	ttg	act	atg	cgt	cgt	atg	cga	gag	tct	gtt	tcc	795	
Pro	Ala	Leu	Pro	Ser	Leu	Thr	Met	Arg	Arg	Met	Arg	Glu	Ser	Val	Ser		
			210					215					220				
agg	atg	cct	gtt	agt	tct	cag	cac	aga	tat	tct	aca	cct	cac	gcc	ttc	843	
Arg	Met	Pro	Val	Ser	Ser	Gln	His	Arg	Tyr	Ser	Thr	Pro	His	Ala	Phe		
		225					230					235					
acc	ttt	aac	acc	tcc	agt	ccc	tca	tct	gaa	ggg	tcc	ctc	tcc	cag	agg	891	
Thr	Phe	Asn	Thr	Ser	Ser	Pro	Ser	Ser	Glu	Gly	Ser	Leu	Ser	Gln	Arg		
	240					245				250							
cag	agg	tcg	aca	tcc	aca	cct	aat	gtc	cac	atg	gtc	agc	acc	acg	ctg	939	
Gln	Arg	Ser	Thr	Ser	Thr	Pro	Asn	Val	His	Met	Val	Ser	Thr	Thr	Leu		
255				260						265					270		
cct	gtg	gac	agc	agg	atg	att	gag	gat	gca	att	cga	agt	cac	agc	gaa	987	
Pro	Val	Asp	Ser	Arg	Met	Ile	Glu	Asp	Ala	Ile	Arg	Ser	His	Ser	Glu		
			275					280						285			
tca	gcc	tca	cct	tca	gcc	ctg	tcc	agt	agc	ccc	aac	aat	ctg	agc	cca	1035	
Ser	Ala	Ser	Pro	Ser	Ala	Leu	Ser	Ser	Ser	Pro	Asn	Asn	Leu	Ser	Pro		
			290					295					300				
aca	ggc	tgg	tca	cag	ccg	aaa	acc	ccc	gtg	cca	gca	caa	aga	gag	cgg	1083	
Thr	Gly	Trp	Ser	Gln	Pro	Lys	Thr	Pro	Val	Pro	Ala	Gln	Arg	Glu	Arg		
		305				310						315					
gca	cca	gta	tct	ggg	acc	cag	gag	aaa	aac	aaa	att	agg	cct	cgt	gga	1131	
Ala	Pro	Val	Ser	Gly	Thr	Gln	Glu	Lys	Asn	Lys	Ile	Arg	Pro	Arg	Gly		
	320					325					330						
cag	aga	gat	tca	agc	tat	tat	tgg	gaa	ata	gaa	gcc	agt	gaa	gtg	atg	1179	
Gln	Arg	Asp	Ser	Ser	Tyr	Tyr	Trp	Glu	Ile	Glu	Ala	Ser	Glu	Val	Met		
335					340					345					350		
ctg	tcc	act	cgg	att	ggg	tca	ggc	tct	ttt	gga	act	gtt	tat	aag	ggg	1227	
Leu	Ser	Thr	Arg	Ile	Gly	Ser	Gly	Ser	Phe	Gly	Thr	Val	Tyr	Lys	Gly		
				355				360						365			
aaa	tgg	cac	gga	gat	gtt	gca	gta	aag	atc	cta	aag	gtt	gtc	gac	cca	1275	
Lys	Trp	His	Gly	Asp	Val	Ala	Val	Lys	Ile	Leu	Lys	Val	Val	Asp	Pro		
			370					375					380				
acc	cca	gag	caa	ttc	cag	gcc	ttc	agg	aat	gag	gtg	gct	gtt	ctg	cgc	1323	
Thr	Pro	Glu	Gln	Phe	Gln	Ala	Phe	Arg	Asn	Glu	Val	Ala	Val	Leu	Arg		
		385				390						395					
aaa	aca	cgg	cat	gtg	aac	att	ctg	ctt	ttc	atg	ggg	tac	atg	aca	aag	1371	

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Lys	Thr	Arg	His	Val	Asn	Ile	Leu	Leu	Phe	Met	Gly	Tyr	Met	Thr	Lys	
400						405					410					
gac	aac	ctg	gca	att	gtg	acc	cag	tgg	tgc	gag	ggc	agc	agc	ctc	tac	1419
Asp	Asn	Leu	Ala	Ile	Val	Thr	Gln	Trp	Cys	Glu	Gly	Ser	Ser	Leu	Tyr	
415					420					425					430	
aaa	cac	ctg	cat	gtc	cag	gag	acc	aag	ttt	cag	atg	ttc	cag	cta	att	1467
Lys	His	Leu	His	Val	Gln	Glu	Thr	Lys	Phe	Gln	Met	Phe	Gln	Leu	Ile	
				435					440					445		
gac	att	gcc	cgg	cag	acg	gct	cag	gga	atg	gac	tat	ttg	cat	gca	aag	1515
Asp	Ile	Ala	Arg	Gln	Thr	Ala	Gln	Gly	Met	Asp	Tyr	Leu	His	Ala	Lys	
			450					455					460			
aac	atc	atc	cat	aga	gac	atg	aaa	tcc	aac	aat	ata	ttt	ctc	cat	gaa	1563
Asn	Ile	Ile	His	Arg	Asp	Met	Lys	Ser	Asn	Asn	Ile	Phe	Leu	His	Glu	
		465					470					475				
ggc	tta	aca	gtg	aaa	att	gga	gat	ttt	ggg	ttg	gca	aca	gta	aag	tca	1611
Gly	Leu	Thr	Val	Lys	Ile	Gly	Asp	Phe	Gly	Leu	Ala	Thr	Val	Lys	Ser	
	480					485					490					
cgc	tgg	agt	ggg	tct	cag	cag	gtt	gaa	caa	cct	act	ggc	tct	gtc	ctc	1659
Arg	Trp	Ser	Gly	Ser	Gln	Gln	Val	Glu	Gln	Pro	Thr	Gly	Ser	Val	Leu	
495					500					505					510	
tgg	atg	gcc	cca	gag	gtg	atc	cga	atg	cag	gat	aac	aac	cca	ttc	agt	1707
Trp	Met	Ala	Pro	Glu	Val	Ile	Arg	Met	Gln	Asp	Asn	Asn	Pro	Phe	Ser	
				515					520					525		
ttc	cag	tgc	gat	gtc	tac	tcc	tat	ggc	atc	gta	ttg	tat	gaa	ctg	atg	1755
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(74) Agent: **VOSSIUS & PARTNER**; Siebertstrasse 4, 81675 München (DE).

(21) International Application Number: PCT/EP01/11282

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(72) Inventors; and

(75) Inventors/Applicants (*for US only*): **HATZOPOULOS, Antonis** [GR/DE]; Siegfriedstrasse 9, 80803 München (DE). **HAUTMANN, Martina** [DE/DE]; Am Hedernfeld 29, 81375 München (DE). **HERBST, Myriam** [DE/DE]; Peter-Henlein-Strasse 23, 81549 München (DE). **GEISHAUSER, Albert** [DE/DE]; Isenerstrasse 7, 85570 Markt Schwaben (DE). **SCHOCH, Jürgen** [DE/DE]; Filchnerstrasse 50, 81476 München (DE).

Published:

— with international search report

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2 October 2003

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: PHARMACEUTICAL COMPOSITIONS COMPRISING POLYNUCLEOTIDES ENCODING A RAF PROTEIN

(57) Abstract: Disclosed are pharmaceutical compositions comprising polynucleotides encoding a Raf protein, vectors, host cells, polypeptides encoded by said polynucleotides as well as agonists or antagonists thereof. Furthermore, described are uses of such pharmaceutical compositions for preventing or treating pathological conditions in which endothelial cells are involved or affected. Finally, methods for screening compounds acting as agonists or antagonists as well as diagnostic compositions and methods are disclosed.



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INTERNATIONAL SEARCH REPORT

Internat Application No
PCT/EP 01/11282

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 A61K38/45

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 A61K C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, MEDLINE, BIOSIS, CHEM ABS Data, SEQUENCE SEARCH

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>US 6 114 517 A (MONIA BRETT P ET AL) 5 September 2000 (2000-09-05)</p> <p>column 1, line 7 - line 53; claims 1-4 column 2, line 24 -column 3, line 4 column 4, line 3 - line 16 column 4, line 39 -column 5, line 17 column 7, line 26 - line 54 column 17, line 1 - line 16</p> <p style="text-align: center;">--- -/--</p>	<p>1,6-14, 20,21, 23,24</p>

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- *&* document member of the same patent family

Date of the actual completion of the international search

20 February 2003

Date of mailing of the international search report

05/03/2003

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INTERNATIONAL SEARCH REPORT

Internat Application No

PCT/EP 01/11282

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	ERHARDT P ET AL: "B-Raf inhibits programmed cell death downstream of cytochrome c release from mitochondria by activating the MEK/Erk pathway." MOLECULAR AND CELLULAR BIOLOGY. UNITED STATES AUG 1999, vol. 19, no. 8, August 1999 (1999-08), pages 5308-5315, XP002231909 ISSN: 0270-7306 page 5308, right-hand column, paragraph 3 page 5312, right-hand column, paragraph 1 -page 5313, right-hand column, paragraph 1 ----	1,2
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X,P	WO 01 12210 A (SCRIPPS RESEARCH INST ;HOOD JOHN (US); CHERESH DAVID A (US); ELICE) 22 February 2001 (2001-02-22) page 1, line 8 - line 25; claims 1-3,19-21,64-67 page 3, line 2 -page 5, line 20 ----	1,6-19
E	EP 1 176 200 A (SWITCH BIOTECH AG) 30 January 2002 (2002-01-30) page 3, line 17 - line 47; claims 2,3,8-10 page 4, line 12 - line 46 ----	1,2,6-8, 20,21, 23,24
E	WO 01 96535 A (SCHEBO BIOTECH AG ;RAPP ULF R (DE); EIGENBRODT ERICH (DE)) 20 December 2001 (2001-12-20) page 1, line 8 - line 15; claims 1-3,5-7,11-13 page 6, line 7 -page 12, line 10 ----	1,6,8, 12,13, 20,21, 23,24
A	WO 97 36587 A (MERCK & CO INC ;HEIMBROOK DAVID C (US); OLIFF ALLEN I (US); STIRDI) 9 October 1997 (1997-10-09) page 1, line 5 - line 31; claims 1-5 page 4, line 25 -page 13, line 23 ----	1,6,8, 12,13
A	SITHANANDAM G ET AL: "B-RAF AND A B-RAF PSEUDOGENE ARE LOCATED ON 7Q IN MAN" ONCOGENE, BASINGSTOKE, HANTS, GB, vol. 7, no. 4, April 1992 (1992-04), pages 795-799, XP008013456 ISSN: 0950-9232 page 798, left-hand column, line 1 ----- -/--	1,6,8, 12,13

INTERNATIONAL SEARCH REPORT

Intern: Application No

PCT/EP 01/11282

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	XU X S ET AL: "A role for c- Raf kinase and Ha-Ras in cytokine-mediated induction of cell adhesion molecules." JOURNAL OF BIOLOGICAL CHEMISTRY, (1998 DEC 11) 273 (50) 33230-8. , XP002231910 page 33231, right-hand column, paragraph 1 page 33238, left-hand column, paragraph 2 - paragraph 3 ---	1,6
A	WO 93 04170 A (US GOVERNMENT) 4 March 1993 (1993-03-04) page 4, line 4 - line 16; claims 1-8 -----	1

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP 01/11282

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☒ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

see FURTHER INFORMATION sheet PCT/ISA/210

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Information on patent family members

Internal application No

PCT/EP 01/11282

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
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